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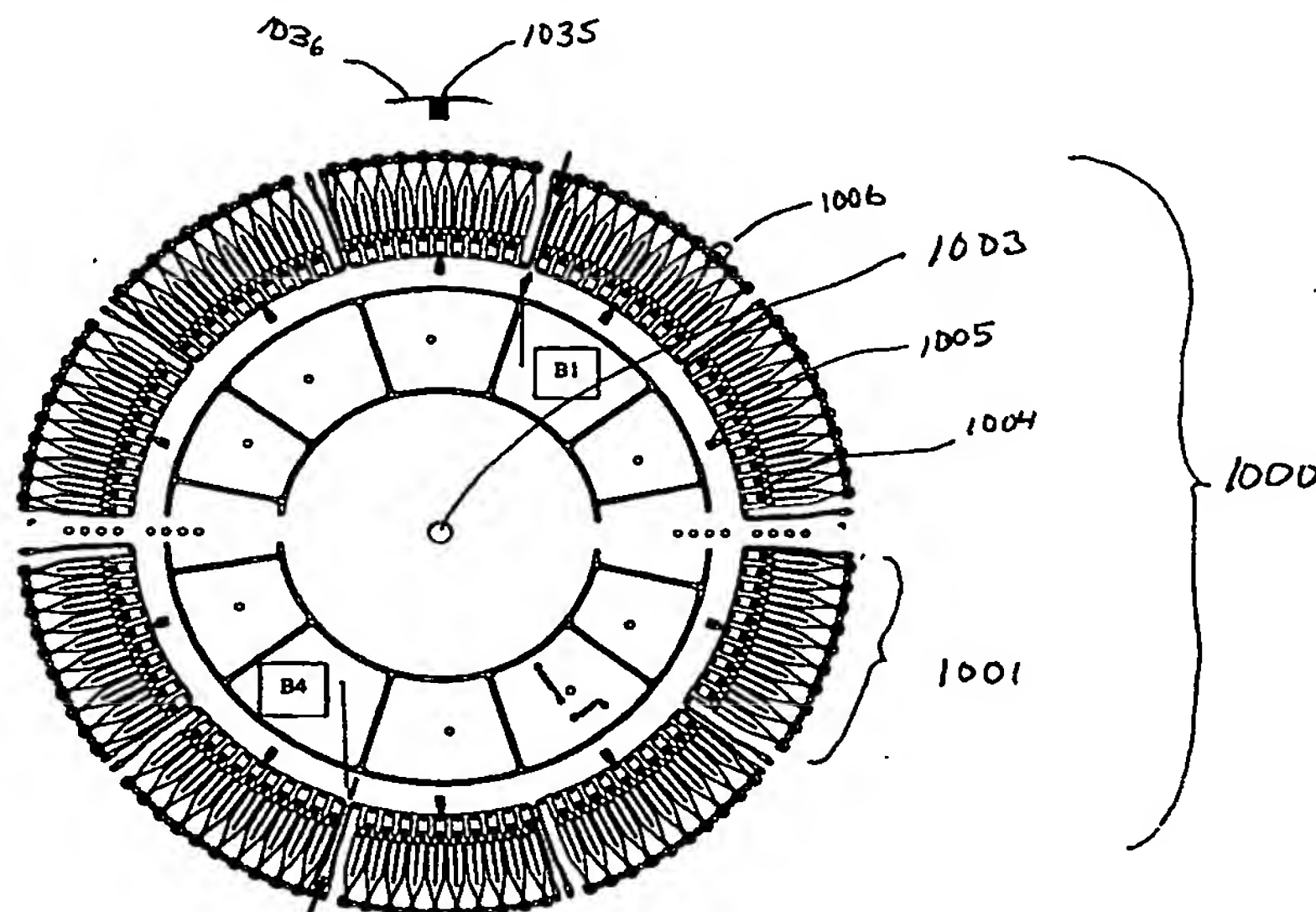
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(54) Title: **A MICROFLUIDIC SYSTEM (MS)**



(57) Abstract: A method for presenting an analyte of a liquid sample as an MS-analyte to a mass spectrometer. The method is characterized in (a) comprising the steps of: (i) applying the liquid sample to a sample inlet port (I) of a microchannel structure (I) of a microfluidic device, said structure also comprising an MS-port, (ii) transporting the analyte by a liquid flow in microchannel structure (I) thereby transforming the analyte to an MS-analyte, and (iii) presenting the MS-analyte to a mass spectrometer via the MS-port, and (b) using inertia force for creating said liquid flow within at least a part of microchannel structure (I). A microfluidic disc comprising (a) an axis of symmetry perpendicular to the plane of the disc, (b) a microchannel structure (I) comprising an inner application area at a shorter radial distance than an outlet port, and an MS-port and a sample inlet port (I).

WO 02/075776 A1

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A MICROFLUIDIC SYSTEM (MS)

TECHNICAL FIELD

The present invention relates to a microfluidic device, which can be interfaced
5 to a mass spectrometer (MS). The device comprises a microchannel structure
having a first port (inlet port) and a second port (outlet port). A sample to be
analysed is applied to the first port and presented to the mass spectrometer in
the second port. This second port will be called an MS-port. There may be
additional inlet and outlet ports. During passage through the microchannel
10 structure the sample is prepared to make it suitable for analysis by mass
spectrometry.

The sample presented in an MS-port will be called an MS-sample. An analyte
in an MS-sample is an MS-analyte. "Sample" and "analyte" without prefix will
15 primarily refer to a sample applied to an inlet port.

Conductive and non-conductive properties are with respect to conducting
electricity.

20 The invention concerns mass spectrometry in which the MS-samples are
subjected to **Energy Desorption/Ionisation** from a surface by input of energy
(EDI MS). Generically this kind of process will be called EDI and the surface
an EDI-surface in the context of the invention. Typically EDIs are thermal
desorption/ionisation (TDI), plasma desorption/ionisation (PDI) and various
25 kinds of irradiation desorption/ionisation (IDI) such as by fast atom
bombardment (FAB), electron impact etc. In the case a laser is used the
principle is called laser desorption/ionisation (LDI). Desorption may be
assisted by presenting the MS analyte together with various helper
substances or functional groups on the surface. Common names are matrix
30 assisted laser desorption/ionisation (MALDI) including surface-enhanced laser
desorption/ionisation (SELDI). For MALDI see the publications discussed
under Background Publications below. For SELDI see WO 0067293
(CIPHERGEN Biosystems).

The invention also concerns electron spray ionisation mass spectrometry (ESI MS).

The term "EDI-area" comprises the EDI-surface as such and the part of a substrate covered by this surface, e.g. the part of the substrate that is under the EDI-surface. Compare the description of figure 5.

The term "microformat" means that in at least a part of a microchannel structure the depth and/or width is in the microformat range, i.e. $< 10^3 \mu\text{m}$, preferably $< 10^2 \mu\text{m}$. The depth and/or width are within these ranges essentially everywhere between an inlet port and an outlet port, e.g. between a sample inlet port and an MS-port. The term "microchannel structures" includes that the channels are enclosed in a substrate.

The term "microfluidic device" means that transport of liquids and various reagents including analytes are transported between different parts within the microchannel structures by a liquid flow.

BACKGROUND PUBLICATIONS.

For some time there has been a demand for microfluidic sample handling and preparation devices with integrated MS-ports. This kind of devices would facilitate automation and parallel experiments, reduce loss of analyte, increase reproducibility and speed etc.

- WO 9704297 (Karger et al) describes a microfluidic device that has an outlet port that is claimed useful when conducting electrospray ionisation mass spectrometry (ESI MS), atmospheric pressure chemical ionisation mass spectrometry (APCI MS), matrix assisted laser desorption/ionisation mass spectrometry (MALDI MS) and a number of other analytical principles.
- US 6,110,343 (Ramsey et al) describe an electrospray interface between a microfluidic device and a mass spectrometer.

- US 5,969,353 (Hsieh) describes an improved interface for electrospray ionization mass spectrometry. The interface is in the form of an electrospray tip connected to a microchannel structure of a chip.
 - US 5,197,185 (Yeung et al) describes a laser-induced vaporisation and ionization interface for directly coupling a microscale liquid based separation process to a mass spectrometer. A light-adsorbing component may be included in the eluting liquid in order to facilitate vaporisation.
 - US 5,705,813 (Apffel et al) and US 5,716,825 (Hancock et al) describe a microfluidic chip containing an MS-port. After processing a sample within the chip the sample will appear in the MS-port. The whole chip is then placed in an MALDI-TOF MS apparatus. The microfluidic device comprises
 - (a) an open ionisation surface that may be used as the probe surface in the vacuum gate of an MALDI-TOF MS apparatus (column 6, lines 53-58 of US 5,705,813) or
 - (b) a pure capture/reaction surface from which the MS-analyte can be transferred to a proper probe surface for MALDI-TOF MS (column 12, lines 13-34, of US 5,716,825).
- These publications suggest that means for transporting the liquid within a microchannel structure of the device are integrated with or connected to the device. The means given are electrical connections, pumps etc. These kinds of transporting means impose an extra complexity on the design and use, which in turn may negatively influence the production costs, easiness of handling etc of these devices.

Although both US 5,705,813 (Apffel et al) and US 5,716,825 (Hancock et al) explicitly concern microfluidic devices, they are scarce about

- the proper fluidics around the MALDI ionisation surface,
- the proper crystallisation on the MALDI ionisation surface,
- the proper geometry of the port in relation to crystallisation, evaporation, the incident laser beam etc,

- the conductive connections to the MALDI ionisation surface for MALDI MS analysis.

These features are important in order to manage with interfacing a microfluidic device to an MALDI mass spectrometer.

5

WO 9704297 (Karger et al) and PCT/SE01/02753 (Gyros AB) suggest a radial or spoke arrangement of the microchannel structures of a microfluidic device.

- 10 WO 9721090 (Mian et al) (page 30, lines 3-4, and page 51, line 10) and WO 0050172 (Burd Mehta) (page 55, line 14) suggest in general terms that their microfluidic systems might be used for preparing samples that are to be analysed by mass spectrometry. WO 9721090 is explicitly related to a system in which centrifugal force is used for driving the liquid flow.

15

A number of publications referring to the use of centrifugal force for moving liquids within microfluidic systems have appeared during the last years. See for instance WO 9721090 (Gamera Bioscience), WO 9807019 (Gamera Bioscience) WO 9853311 (Gamera Bioscience), WO 9955827 (Gyros AB),
20 WO 9958245 (Gyros AB), WO 0025921 (Gyros AB), WO 0040750 (Gyros AB), WO 0056808 (Gyros AB), WO 0062042 (Gyros AB), WO 0102737 (Gyros AB), WO 0146465 (Gyros AB), WO 0147637, (Gyros AB), WO 0154810 (Gyros AB), WO 0147638 (Gyros AB),

- 25 US S.N. 60/315,471 and the corresponding International Patent Application discuss various designs of microfluidic functions, some of which can be applied to the present invention.

See also Zhang et al. "Microfabricated devices for capillary electrophoresis-
30 electrospray mass spectrometry", Anal. Chem. 71 (1999) 3258-3264 and references cited therein.

Kido et al., ("Disc-based immunoassay microarrays", Anal. Chim. Acta 411 (2000) 1-11) has described microspot immunoassays on a compact disc (CD). The authors suggest that a CD could be used as a continuous sample collector for microbore HPLC and subsequent detection for instance by
5 MALDI MS. In a preliminary experiment a piece of a CD manufactured in polycarbonate was covered with gold and spotted with a mixture of peptides and MALDI matrix.

In an International Type Search Report compiled for the priority application
10 US 6,191,418 (Hinsgaul et al), US 4,279,862 (Bretaudiere et al), and US 5,869,830 (Franzen et al) have labelled X/Y. None of these publications concerns problems associated with microfluidic devices and their interfacing with mass spectrometers. US 6,191,418 (Hinsgaul et al) describes a circular arrangement of electrospray tips that can be interfaced one by one to an MS
15 apparatus by rotating the arrangement. The tips are connected to chromatographic columns through which a liquid flow is applied by external means. US 4,279,862 (Bretaudiere et al) describes a circular disc comprising an outwardly directed flow system comprising (a) a unit in which mixing is caused by creating turbulence in a flow which is driven by centrifugal force,
20 and (b) an ending measuring chamber. US 5,869,830 (Franzen et al) describes exact mass determination of MS-analytes that are presented in a conventional way in an MALDI MS apparatus together with a reference compound.

25 OBJECTS OF THE INVENTION.

- A first object is to provide improved means and methods for transporting samples, analytes including fragments and derivatives, reagents etc in microfluidic devices that are capable of being interfaced with a mass spectrometer.
- 30 • A second object is to provide improved microfluidic methods and means for sample handling before presentation of a sample analyte as an MS-analyte. Sub-objects are to provide an efficient concentration, purification and/or transformation of a sample within the microfluidic device while

maintaining a reproducible yield/recovery, and/or minimal loss of precious material.

- A third object is to provide improved microfluidic methods and means that will enable efficient and improved presentation of an MS-sample/MS-analyte. This object in particular applies to MS-samples that are presented on an EDI-surface, or via electrospray ionisation (ESI-tips).
- A fourth object is to enable reproducible mass values from an MS-sample that is presented on an EDI-surface that is present on a microfluidic device in which a liquid flow is caused by inertia force..
- A fifth object is to provide improved microfluidic means and methods for parallel sample treatment before presentation of the MS-analyte to mass spectrometry. The improvements of this object refer to features such as accuracy in concentrating, in chemical transformation, in required time for individual steps and for the total treatment protocol etc. By parallel sample treatment is meant that two or more sample treatments are run in parallel in different microchannel structures within the same microfluidic device. The number of parallel runs may be more than five, such as more than 10, 50, 80, 100, 200, 300 or 400 runs. Particular important numbers of parallel samples are below or equal to the standard number of wells in microtiter plates, e.g. 96 or less, 384 or less, 1536 or less, etc
- A sixth object is to provide a cheap and disposable microfluidic device unit enabling parallel sample treatments and having one or more MS-ports that are adapted to a mass spectrometer.

25 SUMMARY OF THE INVENTION.

The present inventors have recognized that several of the above-mentioned objects can be met in the case inertia force is used for transportation of a liquid within a microfluidic device as defined in this specification. This is applicable to liquid, such as washing liquids and liquids containing at least one of (a) the analyte including derivatives and fragments thereof, (b) a reagent used in the transformation of the sample/analyte, etc.

The present inventors have also recognized that the optimisation of an EDI-area in a microfluidic device is related to

- (a) the design and/or positioning of a conductive layer in the EDI-area, and/or
- (b) the need of a calibrator area associated with an EDI MS-port, and/or
- 5 (c) the need of a proper conductive connection to the EDI-area for MS analysis.

The proper conductive connection will support the proper voltage and/or charge transport at the EDI-area, for instance. Improper conductive properties may negatively affect the mass accuracy, sensitivity, resolution etc. The
10 importance of (a)-(c) increases if there is a plurality of microchannel structures in the microfluidic device.

DETAILED DESCRIPTION OF THE INVENTION

The first aspect of the invention is a method for presenting an analyte of a
15 liquid sample as an MS-analyte to a mass spectrometer. The method is characterized in

- (a) comprising the steps of:
 - (i) applying the liquid sample to a sample inlet port (I) of a microchannel structure (I) of a microfluidic device, said structure also comprising an
20 MS-port,
 - (ii) transporting the analyte by a liquid flow in microchannel structure (I) thereby transforming the analyte to an MS-analyte, and
 - (iii) presenting the MS-analyte to a mass spectrometer via the MS-port, and
- 25 (b) using inertia force for driving said liquid flow within at least a part of microchannel structure (I).

A second aspect of the invention is a microfluidic disc which is characterized in comprising:

- 30 (a) an axis of symmetry perpendicular to the plane of the disc,
- (b) a microchannel structure (I) oriented radially with a liquid flow direction from an inner inlet port towards the periphery of the disc and comprising an MS-port and a sample inlet port (I)

This aspect comprise that the MS-port may be an ESI MS-port or an EDI-MS-port. In case of EDI MS-ports.

The method aspect may also include the mass spectrometric method applied,
5 i.e. the innovative methods may also include the actual collection of a mass spectrum and analysis thereof, for instance in order to gain molecular weight and structure information about an analyte.

The various innovative embodiments of the aspects of the invention are
10 further defined as discussed below.

Liquid flow

The liquid flow used for transport of analyte, analyte-derived entities, reagents etc within the microchannel structures may be driven by electrokinetic forces
15 and/or by non-electrokinetic forces. Typical non-electrokinetic forces are inertia force, such as centrifugal force, capillary forces, forces created by pressure differences etc. For microfluidic devices having circular forms as discussed in this specification it is preferred to drive a liquid flow by spinning the device, i.e. by centrifugal force, in at least a part of each microchannel
20 structures, for instance for the transport into the MS-port. The term "forces created by pressure differences" includes hydrostatic pressure created within certain kinds of microchannel structures by the combined action of spinning and application of a series of liquid aliquots (see below and WO 0146465 (Gyros AB)).

25
At the priority date the most important inertia force to be used in the innovative device and method is centrifugal force, i.e. spinning of the device in order to accomplish an outward transportation of liquid towards the periphery of the disc. The spinning axis coincides with the axis of symmetry of
30 the disc.

Inertia force, such as centrifugal force, may be combined with one or more other kinds of driving forces. The combination may be in the same part of a

microchannel structure. The combination may also mean that inertia force in form of centrifugal force is utilized for transport in a part where the flow shall be directed outwards towards the periphery of a circular disc and other forces in some other part for creating a flow inwards or more or less parallel to the periphery of a disc.

Capillary force may typically be used to transport a liquid aliquot from an inlet port into a microchannel associated with the inlet port. This kind of microchannels may be directed inwards towards the center of a disc or more or less perpendicular thereto.

It may be beneficial to include a pulse giving increased flow for over-coming inter-channel variations in flow resistance, in particular when initiating flow and/or when the liquid is to pass through branchings and curvatures.

15

The sample and its processing.

The sample applied to an inlet port may contain one or more analytes, which may comprise lipid, carbohydrate, nucleic acid and/or peptide structure or any other organic structure. The analyte may also comprise an inorganic structure. The sample treatment protocol to take place within the microchannel structure typically means that the sample is transformed to one or more MS-samples in which

- (a) the MS-analyte is a derivative of the starting analyte and/or
- (b) the amount(s) of non-analyte species have been changed compared to the starting sample, and/or
- (c) the relative occurrence of different MS-analytes in a sample is changed compared to the starting sample, and/or
- (d) the concentration of an MS-analyte is changed relative the corresponding starting analyte in the starting sample, and/or
- (e) sample constituents, such as solvents, have been changed and/or the analyte has been changed from a dissolved form to a solid form, for instance in a co-crystallised form..

Item (a) includes digestion into fragments of various sizes and/or chemical derivatization of an analyte. Digestion may be purely chemical or enzymatic. Derivatization includes so-called mass tagging of either the starting analyte or of a fragment or other derivative formed during a sample treatment protocol, which takes place in the microchannel structure. Items (b) and/or (c) include that the sample analyte has been purified and/or concentrated. Items (a)–(d), in particular, apply to analytes that are biopolymers comprising carbohydrate, nucleic acid and/or peptide structure.

10 The sample is typically in liquid form and may be aqueous.

The sample may also pass through a microchannel structure without being changed. In this case the processing within a microchannel structure only provides a form for dosing of the analyte to the mass spectrometer.

15

FIGURES.

Figure 1-3 illustrate various microchannel structures that have an MS-port.

Figure 4 illustrates an MS-port in form of an electrospray (sideview).

Figures 5a-f illustrate various designs and positions of the conductive layer (I) in MS-ports containing an EDI-surface (cross-sectional sideview of two MS-ports).

Figure 6 illustrates an arrangement around EDI MS-ports with layer (I) and conductive connections (transparent lid, seen from above).

Figures 7a-b illustrate a variant of an EDI-port with a transparent lid (seen from above and in a cross-sectional side-view, respectively).

Figures 8a-b illustrate a variant of microchannel structures suitable to be interfaced with MALDI MS and an optimal arrangement on a full circular microfluidic disc (CD).

The microchannel structures represented in **figures 1-8** are present in planar microfluidic devices.

THE MICROFLUIDIC DEVICE.

The microfluidic device comprises one or more microchannel structures having an inlet port for application of a liquid sample and an MS-port for release and presentation of an MS-analyte to a mass spectrometer. These
5 kinds of ports may coincide in a microchannel structure. There may also be separate inlet ports for application of solvents and reagents and separate outlet ports or waste chambers/cavities for withdrawal of other components that are added and/or produced in the structure. Two or more microchannel structures may have a common inlet port. Depending on the particular design
10 of the device some of the ports may be closed during the sample treatment but opened later on, for instance in order to enable proper release and presentation of the MS-analyte.

The distance between two opposite walls in a microchannel is typically ≤ 1000
15 μm , such as $\leq 100 \mu\text{m}$, or even $\leq 10 \mu\text{m}$, such as $\leq 1 \mu\text{m}$. Functional channel parts (chambers, cavities etc) typical have volumes that are $\leq 500 \mu\text{l}$, such as $\leq 100 \mu\text{l}$ and even $\leq 10 \mu\text{l}$ such as $\leq 1 \mu\text{l}$. In important variants these volumes may be $\leq 500 \text{ nl}$ such as $\leq 100 \text{ nl}$ or $\leq 50 \text{ nl}$. The depths of these parts may be in the interval $\leq 1000 \mu\text{m}$ such as $\leq 100 \mu\text{m}$ such as $\leq 10 \mu\text{m}$ or even ≤ 1
20 μm . The lower limits (width and depth) are always significantly greater than the largest of the reagents and analytes (including fragments and derivatives) that are to be transported within the microchannel structure. The lower limits of the different channel parts are typically in the range $0.1\text{-}0.01 \mu\text{m}$. The aspect ratio (depth to width) may be ≥ 1 or ≤ 1 in all parts or in only a part of a
25 microchannel structure.

Preferred microfluidic devices typically comprise one, two or more, preferably more than 5, microchannel structures. In the preferred variants, the device is formed by covering a substrate surface exposing parts of the microchannel
30 structures with a lid comprising the remaining parts, if any, of the microchannel structures. The lid will prevent or minimise undesired evaporation of liquids as well as facilitate transport of liquids.

A microchannel structure preferably extends in a plane that is common for several microchannel structures. In addition there may be other microchannels that extend in other directions, primarily perpendicular to the common plane. Such other microchannels may function as sample or liquid application areas or connections to microchannel structures that are not located in the common plane, for instance.

The microfluidic devices may be disc-formed and have various geometries, with the circular form being the preferred variant (CD-form). Other variants of discs like the circular form may have an axis of symmetry that is at least 3- or at least 6-numbered. Circular forms typically have radii (r) $\geq 10\%$ or $\leq 300\%$ of the radii of a conventional CD with the conventional CD-format being the preferred.

On devices having circular forms or other forms having an axis of symmetry, an MS-port typically is located at a larger radial distance from the axis of symmetry than an inlet port, a common distribution system/channel etc of a microchannel structure. In case there are more than one inlet ports they may be placed at different radial distances from the axis of symmetry. The flow direction for each microchannel structure is from an inner application area (inlet port, common distribution system or channel etc) towards an outlet port, typically an MS-port, at the periphery of the disc. The microchannel structures may be arranged in the form of one or more concentric circles (annular/circular arrangements) around the axis of symmetry of a disc. The MS-ports in each circle are at the same radial distance from the axis of symmetry.

By the term "radially directed microchannel structure" means that the microchannel structure has an inlet port or a common distribution unit that is closer to the spinning axis (axis of symmetry) than an outlet port, typically the MS-Port. The term does not take into account the design or direction of part structures.

Each microchannel structure may comprise parts that differ with respect to function. In addition to the inlet ports, MS-ports, transportation conduits/channels there may be one or more parts that function as

- 5 (a) application zone/port for reagents and liquids other than sample liquid (second inlet port),
- (b) additional MS-ports,
- (c) reaction zone, for instance for derivatization of an analyte discussed above (digestion, tagging etc).
- (d) pressure creating zone (for instance hydrostatic pressure),
- 10 (e) volume defining zone,
- (f) mixing zone,
- (g) zone for separating and/or concentrating and/or purifying the analyte or a derivative or fragment thereof, for instance by capillary electrophoresis, chromatography and the like,
- 15 (h) waste conduit/chamber/ cavity (for instance in the form of an outlet port),
- (i) zone for splitting a liquid flow, etc.

Each of these parts may have the same or different cross-sectional dimensions as a preceding and/or a subsequent part of the microchannel structure.

20

The sizes of the various parts (a)-(i) depend on number of factors, such as the sample, reagents used, washing, process protocol, desired sensitivity, type of mass spectrometer etc. Typical sizes are found in the range of 1 nl to 1000 μ l, mostly below 1 μ l such as below 500 nl or even below 100 nl such as
25 below 25 or 10 nl (volume defining unit, reactor part, separation part etc). Repeated application of a liquid, e.g. a sample, a washing liquid, a desorption liquid etc to the same inlet port may replace the need for a larger volume defining unit.

30 Splitting of a liquid flow may be located to an upstream part and associated with the inlet so that a starting sample is divided in several aliquots, each of which is then processed in parallel within the device of the invention.

Except for the presence of an MS-port, useful microchannel structures have been described in a number of previous patent publications. See the background publications discussed above.

5 Between parts having different functions there may be valves that can be overcome by increasing the force driving the liquid flow. For variants utilizing spinning, this may for instance be accomplished by increasing the spinning and/or utilizing pressure built up within the structure due to addition of a new portion of liquid combined with spinning. See for instance WO 0040750
10 (Gyros AB) and WO 0146465 (Gyros AB). Valves may be based on capillary junctions (WO 9807019 (Gamera Bioscience)) or hydrophobic breaks (WO 9958245 (Gyros AB), WO 0185602 (Gyros AB & Åmic AB) or on thermic properties of the valve material. The latter kind of valves may be illustrated by so called sacrificing valves (WO 9853311 (Gamera Bioscience)) for instance
15 containing a plug of wax-like material, or reversible valves, for instance containing a thermoreversible polymer in the form of a plug (WO 0102737 (Gyros AB)).

One kind of microchannel structures used according to the invention
20 comprises a zone in which separation and/or concentration and/or purification of the analyte or an analyte-derived entity can take place. This zone is located either before or in the MS-port. Examples of analyte-derived entities are fragments and derivatives of the analyte. This kind of functionality may be particularly important for samples containing low concentrations of analytes,
25 complex mixtures of analytes or high concentrations of interfering substances that may negatively affect the resolution and/or sensitivity of the MS-analysis. The principles utilized for separation, concentration, purification, derivatization, fragmentation etc in the invention are similar to those that are used in the life science area, e.g. separations based on size exclusion and/or
30 on differences in binding to a ligand structure are applicable. Accordingly, a separation zone may contain a separation medium that is capable of binding the analyte or an analyte-derived entity but not the contaminants, or vice versa. The separation medium is typically in particle/bead form, the surface of

the separation zone, or a monolithic plug (porous) that permits through flow. If the analyte or the analyte-derived entity becomes bound, a liquid having the proper desorption characteristics for the bound entity is subsequently allowed to pass through the zone whereupon the bound entity is released and
5 transported downstream. This transport may be directly to the MS-port or to a zone in which a further preparation step is accomplished. Washing steps may be inserted between the sample liquid and the desorption liquid. The separation medium may be soluble or insoluble during the binding step. Soluble separation media are typically insolubilized after binding a desired
10 substance. The principles are well known in the field of macroscopic separations.

Binding as discussed in the preceding paragraph typically means affinity binding or covalent binding to the separation medium. Covalent binding is
15 typically reversible, for instance by thiol-disulfide exchange. Affinity binding (= affinity adsorption) can be illustrated with:

- (a) electrostatic interaction that typically requires that the ligand and the entity to be bound have opposite charges,
- (b) hydrophobic interaction that typically requires that the ligand and the entity
20 to be bound comprises hydrophobic groups,
- (c) electron-donor acceptor interaction that typically requires that the ligand and the entity to be bound have an electron-acceptor and electron-donor group, respectively, or vice versa, and
- (d) bioaffinity binding in which the interaction is of complex nature, typically
25 involving a mixture of different kinds of interactions and/or groups.

Ion exchange ligands may be cationic (= anion exchange ligands) or anionic (= cation exchange ligands). Typical anion exchange ligands have positively charged nitrogen, the most common ones being primary, secondary, tertiary
30 or quarternary ammonium ligands, and certain amidinium groups. Typical cation exchange ligands have negatively charged carboxylate groups, phosphate groups, phosphonate groups, sulphate groups and sulphonate groups.

Bioaffinity binding includes that the analyte or the analyte-derived entity is a member of a so-called bioaffinity pair and the ligand is the other member of the pair. Typical bioaffinity pairs are antigen/hapten and an antibody/antigen binding fragment of the antibody; complementary nucleic acids; immunoglobulin-binding protein and immunoglobulin (for instance IgG or an Fc-part thereof and protein A or G), lectin and the corresponding carbohydrate, etc. The term "bioaffinity pair" includes affinity pairs in which one or both of the members are synthetic, for instance mimicking a native member of a bioaffinity pair.

10

If the analyte in a sample has peptide structure or nucleic acid structure or in other ways has a pronounced hydrophobicity, the separation medium may be of the reverse phase type (hydrophobic) combined with using desorption liquids (eluent) that are organic, for instance acetonitrile, isopropanol, methanol, and the like. Depending on the particular sample and the presence of analytes or analyte-derived entities, which have a common binding structure, a group-specific separation medium may be utilized. The separation medium may thus, like a reverse phase adsorbent, result in an MS-sample that has a reduced concentration of salt, i.e. in desalting.

20

In each microchannel structure there may be two or more separation zones utilizing the same or different principles such as size and charge. For amphoteric substances such as proteins and peptides the latter principle may be illustrated with isoelectric focusing.

25

After a separation step comprising binding to a separation medium the concentration of an analyte or an analyte-derived entity in the desorption liquid after passage of the separation medium is typically higher than in the starting sample. The increase may be with a factor $> 10^0$, for instance in the interval 10^1 - 10^6 , such as 10^1 - 10^4 .

30

As already mentioned a separation zone may be combined with zones for derivatization including fragmentation. There may also be microchannel structures that have a derivatization zone but no separation zone.

- 5 U.S S.N. 60/322,621 and the corresponding International Application describes the use of the above-mentioned affinity principles in an assay without explicitly referring to mass spectrometry.

Figure 1 illustrates a microchannel structure that comprises (a) an inlet port
10 (1) for liquids including the sample liquid, (b) an MS-port (2) comprising for instance an EDI-surface, (c) a flow conduit (3) between the inlet port (1) and the MS-port (2). The MS-port may be open or covered. The flow conduit (3) may have a zone (4) containing an adsorbent for separation/concentration. If there are several microchannel structures in a device there may be a
15 common application area/channel with openings for the inlet ports (not shown). The MS-port may be an EDI MS-port, an eletrospray MS-port.

The structure of **figure 1** may be present on a circular disc with the inlet port (1) closer to the centre than the MS-port (2). If the MS-port is an EDI-MS port
20 and liquid is transported through the conduit (3) by spinning the disc, liquid will leave the MS-port either as drops or by evaporation depending on the vapour pressure of the liquid and/or the spinning speed. A lower vapour pressure and an increased spinning speed will promote drop formation while a higher vapour pressure and a decreased spinning speed will promote
25 evaporation of the liquid and crystallisation of the MS-analyte in the mS-port. A too low spinning speed and a too low vapour pressure will increase the risk for deposition of material in the conduit (3).

Figure 2 illustrates another variant of a microchannel structure. It has two
30 inlet ports (5,6) that may be used for application of sample, washing liquid and desorption liquid. One of the inlet ports (5) is connected to an application area/channel (7) that may be common to several microchannel structures in the same device. This first inlet port (5) is connected to one of the shanks (8)

of a U-shaped channel via the application area/channel (7). The other inlet port (6) is connected to the other shank of the U. In the lower part of the U there is an exit conduit (9) leading to an MS-port (10). The exit conduit (9) may comprise a zone (12) containing a separation medium. From the MS-port (10) there may be a waste channel (13) leading to a waste channel (14) that may be common for several microchannel structures in the same device. Conduit (9) may comprise a valve function, for instance in the form of a hydrophobic break, upstream a possible separation zone (12).

10 The microchannel structure of **figure 2** is also adapted to a circular disc and driving liquid flow by spinning the disc. The application channel (7) is at a shorter radial distance from the centre of the disc than waste channel (14).

Figure 3 illustrates a microchannel structure which comprises a separate sample inlet port (14), an MS-port (15) and therebetween a structure that may be used for sample preparation. In this variant there is a volume-defining unit comprising a metering microcavity (16) between the sample inlet port (14) and MS-port (15) with an over-flow conduit (17) that ends in a waste chamber (25a) that may be common for several microchannel structures. At the lower part of the metering microcavity (16) there is a first exit conduit (18) leading to one of the shanks (19) of a U-shaped channel. The other shank (20) of this U may be connected to an inlet port (21) for washing and/or desorption liquids. At the lower part of the U-shaped channel there may be a second exit conduit (22) leading into one of the shanks (23) of a second U-shaped channel. The other shank (24) may be connected to a waste channel (25b) that after a bent (26) may end in a waste chamber (25a). At the lower part of the second U-formed channel there may be a third exit conduit (27) leading into the MS-port (15) that may contain an EDI-surface or an electrospray unit. In order to control the flow in the microchannel structure, valve functions may be located (a) in the first exit conduit (18), for instance immediately downstream the volume-defining unit (16), (b) possibly also in the second exit conduit (22), for instance immediately after the first U,

(c) in the third exit conduit (27), for instance immediately after the second U,
and

(d) in association with the connection between the overflow channel (17) and
the waste channel (25b).

5 The valves may be of the types discussed above with preference for hydrophobic breaks. A suitable adsorbent (28) as discussed above may be placed in the second exit conduit (23) and may also function as a valve. In the case the adsorbent is in the form of particles they are preferably kept in place by a constriction of the inner walls of the conduits.

10

The structure presented in **figure 3** is adapted for transporting liquid with centrifugal forces, i.e. with the structure present in a disc and oriented radially outwards from the centre of the disc. At the start of an experiment the metering cavity (16) is filled up with sample liquid at least to the connection
15 between the over-flow channel (17) and the metering cavity (16), for instance by capillary action. Liquid will enter the overflow channel (17). By first overcoming the valve function between the overflow channel (17) and the waste channel (25a), excess liquid will pass into the waste channel (25a). By then overcoming the valve function in the first exit conduit (18), the liquid in
20 the metering microcavity (16) will pass into the first U and down through the adsorbent (28) where the analytes are captured. The liquid now being essentially devoid of analyte will then halt at the bottom of the second U. In the next step, one or more aliquots of a washing liquid may be applied through either of the inlet ports (14,21), i.e. through the second shank (20) of
25 the first U or via the same inlet port (14) as the sample. A washing liquid will pass through the adsorbent (28), collect in the bottom of the second U and push the liquid already present into the waste chamber/channel (25a,b). Subsequently, a desorption liquid is applied through either of the two inlet ports (14,21) and passed through the adsorbent (18) where it releases the
30 analyte and into the bottom of the second U where it pushes the washing liquid into the waste chamber/channel (25a,b). The desorption liquid containing released analyte is then passed into the MS-port (15) from the

bottom of the second U by overcoming the valve function in the third exit conduit (27).

The operations are preferably carried out while spinning the disc. If the valves
5 are in the form of hydrophobic breaks they can be passed by properly
adapting the g-forces, i.e. by the spinning. By properly balancing the
hydrophilicity/hydrophobicity of a liquid, passage or non-passage through a
valve may be controlled without changing the spinning speed. This is
illustrated by utilizing a hydrophobic break as the valve in the third exit conduit
10 (27) combined with utilizing water-solutions as samples and washing liquids
and liquids containing organic solvents as desorption liquids. In the
alternative, valves that are opened by external means can be used. By
placing the outlet of the first exit conduit (18) at a shorter radial distance from
the axis of symmetry than the lowest part of the metering microcavity (16)
15 particulate matters, if present in the sample, will sediment and be retained in
the volume-defining unit when the metering microcavity (16) is emptied
through the first exit conduit (18).

Calibrator areas (29) are shown in each of **figures 1-3**. Each calibrator area
20 may be connected to a common area for application of a calibrator
substance.

These kind of flow systems has been described in WO 0040750 (Gyros AB)
and WO 0146465 (Gyros AB) which are hereby incorporated by reference.

25

In certain variants the inlet port for the sample and the MS-port may coincide.
In this case the MS-port preferably comprises the surface on which the
analyte can be collected (adsorbed). Remaining liquid and washing liquids, if
used, are passed into the microchannel structure that then will function as a
30 waste channel and possibly contain a separate outlet port particularly adapted
for wastes and the like, or a waste chamber. In order to accomplish a
concentrating and/or separating effect the surface may expose structures
selectively binding/capturing the analyte as discussed above for a separation

zone. This variant also encompasses that there may be a separate inlet port for washing and desorption liquids and microchannel part communicating with the combined sample and MS-port.

5 The MS-port

The MS-port typically has an conductive part. The conductive part may for instance be present in an EDI area or in a nozzle suitable for electrospray ionisation, for instance a nanospray, or in any other form device that is used to present a sample to a mass spectrometer. An electrospray nozzle provides
10 an orifice for instance in the form of a tip with a through-passing hole. Various kinds of sample presentation devices have been described in the publications discussed above.

There may be a valve in the microchannel before its inlet to the MS-port.

15

The term conductive material includes semi-conductive material, although materials having a conductivity that is larger than silicon or larger than germanium are preferred. A typical conductive material comprise:

- (a) metals such as copper, gold, platinum etc, mixtures of metals (alloys),
20 such as stainless steel etc,
- (b) conductive metal oxides and mixtures thereof, such as indium oxide, tin oxide, indium tin oxide etc,
- (c) conductive polymers which includes polymers that are conductive as such and conductive composites containing a non-conductive polymer and a
25 conductive material, for instance according to a)-c) and other conductive composites, etc.

ESI MS-ports

Figure 4 illustrates an MS-port suitable for electrospray ionisation in a mass
30 spectrometer. This kind of port may be located where an MS-port has been indicated in any of the structures given in **figures 1-3**. The MS-analyte may thus be collected in an MS-port comprising a collection zone (30), which zone is in fluid communication via the electrospray conduit (31) with the outlet

orifice (32). The electrospray conduit may be in the form of a tip. The MS-analyte is entering the MS-port via conduit (33). The orifices (32) of the electrospray arrangement are preferably positioned on the edge of a disc with one, two or more orifices per microchannel structure. Typical disc-forms have
5 been discussed above. In use an electrospray orifice is matched to the sampling orifice of a mass spectrometer and liquid in the collection zone (30) is sprayed into the mass spectrometer. In a preferred variant the disc is circular. The arrangement of the electrospray tips is preferably annular around the centre of the disc. The orifices are preferably located at the edge
10 of the disc with a radial spray direction. The electrospray orifices may alternatively be in one planar side of the microfluidic device with a spray direction having a component that is perpendicular to the side. Annular arrangements preferably at the edge of a circular disc will simplify accurate positioning of the electrospray orifices relative to the sample application
15 opening of a mass spectrometer.

Electrospray units suitable for electrospray ionization mass spectrometry (ESI MS), for instance adapted to the nanospray format, are mostly formed in capillaries made of glass or fused silica, or polymer material like silicon. The
20 tubings are typically of cylindrical geometry with tip internal diameters in the 5-20 μm range. The word nanospray means that the liquid transferred out of the tubing is in the nanoliter per minute range. Suitable rates for transfer of liquid to the mass spectrometer can be found in the interval of 1-1000 nl/min, e.g. in the interval 10-500 nl/min. By infusion (no external force), only a few
25 nanoliters per minute (5-25 nl/min) is transported out of the tubing while with applied external pressure 50-500 nl/min is more common.

A liquid solution suitable for ESI MS analysis comprises an organic solvent:water mixture and includes a lower concentration of acid or base. The
30 composition is important especially with regard to surface tension and conductivity. A low surface tension and a low conductivity are desirable in order to obtain an efficient desolvation and ionization process and a stable spray. If the sample is dissolved in water only, a so-called make-up solvent is

preferably added (external delivery). A make-up solvent is typically configured co-axially (sheath-flow) around the nanospray tip. A make-up gas (typical N₂) may be added (e.g., co-axially) to assist the desolvation process. Creation of a suitable liquid composition of the MS-sample may be part of the sample
5 preparation process taking place upstream the MS port in other parts of microfluidic device.

The tip geometry is important for a stable spray. Preferably the tip is pulled from a cylindrical tubing which result in an oblong tip with a conical shape.
10 The outer diameter of the tubing near the orifice then becomes of similar dimension as the internal diameter.

In order to induce a spray from an electrospray type (towards the inlet of the mass spectrometer) a voltage has to be applied on the tip. Therefore the tip
15 has to be made conductive. Different kind of metals can be deposited by different techniques onto the tip (or part of the tubing). Important aspects here regard the stability (life-time) of the metallized tip since the voltage applied as well as different solvents affects its stability. Other possibilities also exist, e.g., an electrode can be inserted into the tip whereby a voltage can be applied to
20 induce electrospray. Another alternative is to make the tip in a material comprising any of the above-mentioned conductive materials. Typical voltages used in nanospray range between 500-2000 volts.

Typical electrospray nozzles are available from a number of manufacturers,
25 for instance New Objective, MA, U.S.A. A variant that is believed to have advantages for microfluidic devices is presented in PCT/SE01/02753 (Gyros AB). See also WO 9704297 (Karger et al), US 5,969,353 (Hsieh) and US 6,110,343 (Ramsey et al) discussed above.

30 **EDI MS-ports**

The MS-port may also be used for EDI-MS and will then contain an EDI-area.

Upward and downward directions when used in the context of EDI areas refer to the directions defined in the figures irrespective of how the area is positioned in a mass spectrometer.

- 5 EDI MS ports may be adapted to different EDI mass spectrometry variants, for instance Time of Flight (TOF), Quadropole, Fourier-Transformed Ion Cyclotron Resonance (FT-ICR), ion trap etc.

10 The EDI MS-port requires a free passage for the release of the ions created during desorption/ionisation and thus has an opening straight above the EDI-surface. This opening should be coaxial with and cover the EDI-surface. In other words the EDI MS port is typically in form of a well or depression with the EDI-surface at the bottom and in fluid communication at least with upstream parts of the corresponding microchannel structure. This includes
15 that the opening may be covered during the sample treatment within the microfluidic device but subsequently opened to enable desorption/ionisation and possibly also evaporation of solvents. If an IDI principle is used the opening should also provide space for the incident irradiation.

- 20 An EDI-surface may in principle have any geometric form although preferred forms should be as compact as possible, for instance regular forms, such as squares and square-like forms, and rounded forms, such as circular and circle-like forms. The size of an EDI-surface preferably is the same as a circle with a diameter in the interval of 25-2000 μm . There may be advantages if the
25 cross-sectional area of the incident beam used for irradiation is able to encompass the complete EDI-surface or as much as possible, for instance more than 25 % or more than 50 %.

An EDI-area comprises a conductive layer (layer I), for instance a metal layer
30 of copper, gold, platinum, stainless steel etc or a layer of any other conductive material of the kinds discussed above. Layer (I) may coincide with the EDI-surface or be parallel thereto. Layer (I) has a conductive connection for supporting the proper voltage and charge transport at the EDI-surface. The

complete EDI-area from the lowest part to the EDI-surface may be made of conductive material, i.e. correspond to layer (I). In the case the microfluidic device comprises more than one microchannel structure with an EDI MS-port, layer (I) of one EDI MS-port may be part of a common continuous conductive layer which extends into and encompasses layer (I) of two or more of the other EDI MS-ports. In preferred variants the common continuous layer comprises layer (I) of all EDI-MS ports of a microfluidic device. The common conductive layer may be essentially planar. The common conductive layer may have depressions corresponding to the EDI-surfaces and/or to other parts of the microchannel structures of the innovative device. Typical variants are that the common conductive layer is positioned

- (a) on top of the microfluidic device or
- (b) between two substrates that are joined together to form the enclosed microchannel structures of a microfluidic device.

In both variants the common conductive layer extends into the inner walls and layer (I) of the MS-ports. The MS-ports correspond to depressions.

The exact geometric shape of layer (I) outside the MS-port depends on the particular device and practical ways of its manufacture. For instance a common conductive layer may have an annular or arc-like form in case the MS-ports are annularly arranged.

In one innovative variant, the EDI-area comprises a non-conductive layer (layer (II)), which covers the conductive layer (I). Layer (II) in one EDI-area may extend into and encompass layer (II) in two or more of the other EDI-areas as described for layer (I).

In another innovative variant the device has a separate conductive layer (III) positioned above the common plane defined by the surface of each EDI-area of a device and not connected to layer (I) in different EDI MS-ports. Layer (III) has openings matching the EDI-surfaces and permitting irradiation of these surfaces and escape of ions through the openings.

These innovative variants of EDI-areas are schematically illustrated in **figures 5a-f**, each of which shows a cross-sectional view across the EDI-areas of two MS-ports in a microfluidic device according to the invention. The EDI-surfaces are referenced as (51) and the EDI-areas as (52) (within the dotted vertical lines, a and b in figure 5f). Each MS-port comprises the EDI-area plus the corresponding depression. The conductive layers (54) are hatched. It is apparent that each EDI-area comprises a conductive layer (I) (53).

Figure 5a shows a common continuous conductive layer (54) at the bottom of the device which layer encompasses layer (I) (53) of each EDI-area (52). A non-conductive layer (II) (55) is placed between layer (I) (53) and the EDI-surface (51). **Figure 5b** shows a variant, which is similar to the variant in figure 5a, but the common continuous conductive layer is embedded within the device. Non-conductive layer (II) (55) is present. In **figure 5c** there is a common continuous conductive layer (54) comprising the EDI-surfaces and layer (I). In **figure 5d** there is no common continuous conductive layer. Layer (I) (53) for different MS-ports are isolated from each other and correspond to EDI-surfaces. **Figure 5e** shows a variant in which there is a separate continuous conductive layer (54) above layer (I) (53) of the EDI-areas. This conductive layer (54) has openings (56) corresponding to the openings of each MS-port and may be a surface layer on the upper or lower side of a lid covering the microchannel structures. **Figure 5f** shows a variant in which there is a common continuous conductive layer comprising layer (I). The EDI-surfaces coincides with layer (I) in the MS-ports. The continuous layer also encompasses the inner walls of the MS-ports. The MS-ports appear as depressions in the common conductive layer.

For variants in which the open microchannel structures have been fabricated in a base substrate and covered by a lid, the base substrate may consist of conductive material and correspond to layer (I). In these variants the lid may comprise a non-conductive or conductive material.

Figure 6 illustrates an arrangement of MS-ports on a circular disc (with a transparent lid), in which layer (I) (34) of each MS-port has a connection for conductivity (35) with a peripheral conductive layer (36) which is closer to the edge of the disc than the MS-ports. In this variant each microchannel structure (37) comprises an MS-port and extends upstream to an inlet port (38). Layer (I) (34), the connections (35) and layer (36) may be interpreted as a continuous conductive layer.

Figures 7a-b illustrate an MS-port in which the opening above an EDI-surface is defined by a hole (39) in a lid (40) which in this case is transparent. The incoming microchannel (41) opens to a circular area (42) with a diameter, which is less than the diameter of the hole (39). Layer (I) (43), EDI-area (44), EDI-surface (45) are between the two dotted lines. Layer (I) extends into a common conductive layer (46). This design in which the MS-port provides an opening, which is greater than the EDI-area will facilitate for an incident beam to cover any spot of the EDI-surface. In preferred variants the microchannel (41) extends into the bottom of the MS-port as an open microchannel of constant depth. Seen from above the microchannel may be widening like an expanding droplet.

20

A conductive layer per se may function as a conductive connection or there may be distinct connections (35) to layer (I). See **figure 6**.

In certain variants the lid that covers the microchannel structures also covers the EDI-surfaces. For these variants the lid is removable at least at the MS-ports. After processing of a sample in an upstream part of a microchannel structure and transportation of the treated sample to the covered MS-port, the lid is removed thereby permitting evaporation of solvents from the MS-port and irradiation in order to accomplish desorption/ionisation of MS-analyte molecules.

30

Liquids entering the MS-ports.

During transport through a microchannel structure the solvent composition may be changed to fit the particular kind of mass spectrometer used. In the case of microchannel structures comprising EDI MS-ports and separation zones containing a separation medium, a compound (= EDI-matrix) that upon
5 co-crystallisation with the analyte or analyte-derived entity assists desorption/ionisation may be (a) included in the desorption liquid, (b) included in another liquid that is also guided to the MS-port, or (c) predispensed to the EDI-surface or dispensed to this surface after the analyte or analyte-derived entity has been deposited on the EDI-surface. There may also be included
10 compounds that facilitate crystallization on the EDI-surface. Both kinds of helper compounds may be included even there is no separation zone.

Calibration of the mass scale.

To ensure accurate mass determination, calibrator areas (spots) containing a
15 compound of known molecular weight (standard, calibrator substance) may be present in the proximity of an MS-port. Calibrator areas (29) are shown in figures 1-3. Alternatively, the standards may be included in the sample or added to an EDI-area before desorption/ionisation (internal calibrator). The choice of calibrator substance, its amount etc will depend on its use as an
20 external or internal calibrator, the MS-analyte and its concentration etc.

Material from which the microfluidic device is manufactured.

The microchannel structures are typically fabricated in inorganic and/or organic material, preferably plastics or other organic polymers. The material
25 may be conductive or non-conductive as already discussed. Certain parts of a microchannel structure may be metalized.

Suitable organic polymers may derive from polymerisation of monomers comprising unsaturation, such as carbon-carbon double bonds and/or carbon-
30 carbon-triple bonds. The monomers may, for instance, be selected from mono-, di and poly/oligo-unsaturated compounds, e.g. vinyl compounds and other compounds containing unsaturation.

Another type of organic polymers that may be used is based on condensation polymers in which the monomers are selected from compounds exhibiting two or more groups selected among amino, hydroxy, carboxy etc groups. The plastics contemplated are typically polycarbonates, polyamides, polyamines, 5 polyethers etc. Polyethers include the corresponding silicon analogues, such as silicone rubber.

The polymers are preferably in cross-linked form.

10 The plastics may be a mixture of two or more different polymer(s)/copolymer(s).

At least a part of the microchannel structure may have a surface that has been derivatised and/or hydrophilized, for instance by being coated with a 15 non-ionic hydrophilic polymer according to the principles outlined in WO 0147637 (Gyros AB) or by treatment in gas plasma. Typical gas plasma treatments utilize non-polymerisable gases, for instance as outlined in WO 0056808 (Gyros AB). A hydrophilized surface may also be functionalized in order to introduce one or more functional groups that are capable of 20 interacting with the sample analyte, an analyte-derived compound or one or more of the reagents added. Surfaces may be made of copper, gold, platinum, stainless less etc, for instance by metallization, in order to enable a desired derivatization or for providing a conductive surface, for instance in an MS-port. Gold surfaces for instance may be derivatized by reaction with thiol- 25 containing compounds that have a desired functionality, for instance hydrophilicity.

The optimal water contact angle for the surfaces within a structure depends on the protocols to be carried out, the dimensions of the microchannels and 30 chambers, composition and surface tension of the liquids, etc. As a rule of thumb, the surface of one, two, three or four of the inner walls (side-walls, bottom or top), of a microchannel in a microfluidic device have to be wettable the liquid used, preferably aqueous liquids, such as water. Preferred water

contact angles are $\leq 40^\circ$ or $\leq 30^\circ$, such as $\leq 25^\circ$ or $\leq 20^\circ$. These figures refer to values obtained at the temperature of use, primarily room temperature.

It is believed that the preferred variants of the inventive microfluidic devices
5 will be delivered to the customer in a dry state. The surfaces of the microchannel structures of the device therefore should have a hydrophilicity sufficient to permit the aqueous liquid to be used to penetrate different parts of the channels of the structure by capillary forces (self-suction). This of course only applies if a valve function at the entrance of the particular part
10 has been overcome.

Best Mode

The best mass spectrometric results accomplished at the priority date have been obtained for the variant described in example 4 below.

15

The best mode at the filing date is illustrated by example 5.

The invention is further defined in the appending claim and will now be illustrated with a non-limiting experimental part.

20

The following patents and patent applications have been referenced in this specification and hereby incorporated by reference:

WO 9116966 (Pharmacia Biotech AB), WO 9704297 (Karger et al), WO
9721090 (Gamera Bioscience), WO 9807019 (Gamera Bioscience) WO
25 9853311 (Gamera Bioscience), WO 9955827 (Gyros AB), WO 9958245
(Gyros AB), WO 0025921 (Gyros AB), WO 0040750 (Gyros AB), WO
0056808 (Gyros AB), WO 0062042 (Gyros AB), WO 0102737 (Gyros AB),
WO 0146465 (Gyros AB), WO 0147637, (Gyros AB), WO 0154810 (Gyros
AB), WO 0147638 (Gyros AB), WO 0185602 (Åmic AB & Gyros AB),
30 PCT/SE01/022753 (Gyros AB & Åmic), US 5969353 (Hsie), and US
6,110,343 (Ramsey et al), and U.S. S.N. 60/315,471, U.S. S.N. 60/322,621
and corresponding International Applications.

EXPERIMENTAL PART**EXAMPLE 1. Gold at different positions in a CD**

Gold patterning	Sensitivity*	Charging of substrate**
No gold	Poor	Yes
Gold on all sides	Good	No
Gold on upper side	Good	No
Gold on bottom side	Good	Yes
Isolated gold spots on the upper side	Good	Yes
Gold spots on the upper side. Every spot being conductively connected contact with the adapter plate through an individual gold string or a common gold area.	Good	No

5

* Good = sensitivity for an in-solution tryptic digest of BSA comparable to the sensitivity obtained on a conventional stainless steel target

** Charging is observed as significant mass shift (≥ 1 Da) upon repeated laser desorption/ionization and/or loss of signal.

10

This table shows the results form a summary of experiments performed before the priority date in order to optimise the design of the CD-MALDI interface. Gold was sputtered at various positions of the CD and the MALDI characteristics were studied for a tryptic digest of Bovine Serum Albumin (BSA). The CD was placed on a metal adapter inserted into the ion source. The gold was hence patterned in various ways to determine the importance of electrical contact between the MALDI ports and the adapter plate.

15

EXAMPLE 2. Planar CD and structured removable lid.

This example shows a planar CD in combination with a lid in which the microfluidic structures are present. The structured lid was achieved through casting Memosil (Hereaus, Germany) against a nickel-coated master. The
5 microfluidic structure employed in this example is shown in **figure 2**.

The structured lid is attached to the CD by adhesion forces. The surface facing the lid should be hydrophilic as the presented invention utilizes capillary action to fill the microfluidic structures. This is especially important as
10 the moulded lid, being a type of silicon rubber is hydrophobic.

The upper side of the CD was covered with gold using a DC Bias magnetron sputtering method (1×10^{-5} torr, Ar plasma and titan as adhesion layer) and made hydrophilic according to the following procedure; The gold sputtered
15 side was cleaned by rinsing with ethanol, followed by an oxygen plasma treatment (Plasma Science PSO500,). After plasma cleaning a self-assembled monolayer (SAM) of hydroxylthiol was formed on the gold surface. The hydroxylthiol was 11-mercapto-1-undecanol (Aldrich, Milwaukee, WI) and used at a concentration of 2mM in degassed ethanol. To obtain a well-
20 organized SAM, the gold sputtered disc was immersed in the thiol solution over night. After the thiol adsorption the CD was sonicated in ethanol for ca 2 min.

The lid, containing the microfluidic channels, was attached to the CD by
25 adhesion forces. A second piece of polymeric material was mounted at a position of 180° from the structured lid as a counterbalance. Reversed phase beads (Source 15 RPC, Amersham Pharmacia Biotech, Sweden) with a diameter of $15 \mu\text{m}$ were packed into the individual structures using the filling port present in the common distribution channel. The slurry, containing the
30 beads, was drawn into the individual channels by capillary action. Eighteen parallel reversed phase columns were formed when the disc was spun at 3000 rpm for 1 minute. The columns were rinsed with water containing 0.1% TFA (trifluoroacetic acid, Aldrich)) two times. The rinsing was performed at an

rpm of 2500 for ca 1 min. 200 nL of in-solution tryptic digest of BSA was added to individual channels through the sample inlet. The following procedure was used for digestion. The BSA (Sigma) was dissolved to a final concentration of 4.75 pmol/ μ L in 0.1 ammonium bicarbonate buffer at pH8.

- 5 The enzyme-modified trypsin (Promega Corp., Madison, WI) was added and dissolved at a ratio of BSA/trypsin 20:1. The sample was incubated at 37°C for 4 hours and then stored at -20 °C until used.

The sample was allowed to pass over the reversed phase columns at 1500
10 rpm. A second rinsing/washing step was performed as above using water containing 0.1% TFA. Finally the peptides were eluted using 200 nL eluent consisting 50% isopropanol, 50 % water and α -cyano-4-hydroxycinnamic acid (Aldrich) below saturation. The eluent was prepared by saturating a
15 100 μ L of this mixture 200 μ L of 50% water:50% isopropanol was added, resulting in an eluent saturated to approximately 2/3 with α -cyano-4-hydroxycinnamic acid.

The presentation of the sample in the MALDI port was performed in two
20 different ways.

- a) In the first example a full structure was utilised (**figure 2**). Eluent from the column was collected in the container placed at an outer radial position relative of the reversed phase column. When the lid was removed the liquid quickly evaporated leaving co-crystallized matrix
25 and peptides on the gold sputtered surface. The disc was cut in halves to fit in the MALDI ionisation interface.
- b) The moulded structure was cut directly after the packed column leaving an open-ended microstructure. The eluent was allowed to pass the column at a pre-determined speed (1500 rpm) in order to generate a
30 controlled evaporation of the solvent at the MALDI port, and hence the formation of co-crystallized matrix and peptides suitable for MALDI

analysis. The disc was cut in halves to fit in the MALDI ionisation interface.

EXAMPLE 3. Structured CD and site-specific elution

- 5 This example employs a CD with integrated microfluidic structures, a thin (≤ 70 μm) lid with holes at positions matching the MALDI port in the CD. The microfluidic structure employed in this example is shown in **figure 1**.

The polycarbonate CD was covered with gold as described above. The side
10 was hydrophilized using the thiolprocedure described above. The lid (SkultunaFlexible, Skultuna, Sweden), having, pre-drilled holes, was attached to the CD through heat pressing at 135°C.

Reversed phase beads (Source 15 RPC) with a diameter of 15 μm were
15 packed in the individual structures using capillary forces in combination with centrifugation. The columns were rinsed with ethanol and spun to dryness before 23 fmol of tryptically digested BSA was added and spun down using 700 rpm. The tryptic digest of BSA was generated according to the procedure described above. After sample addition, the column was rinsed
20 twice with water. α -cyano-4-hydroxycinnamic acid was mixed in an organic solvent of acetonitrile/water 3:7 containing 0,1% TFA to a saturation of 2/3 and 250 nl was used to elute the sample from the 3 nl packed column.

The crystals obtained after evaporation of the organic/water mixture
25 contained co-crystallized peptides. Eight singly charged peptide peaks were present in the mass spectrum obtained.

EXAMPLE 4. Parallel sample preparation in a product CD.

Description of the microfluidic disc (CD)

- 30 **Figure 8a** illustrate a product microfluidic device (CD) (1000) comprising 10 sets (1001) of identical microchannel structures (1002) arranged annularly around the spinning axis (axis of symmetry) (1003) of a circular disc (1000). Each set comprises 10 microchannel structures. Each microchannel structure

is oriented radially with an inlet port (1004,1005) located at shorter radial distance than an outlet port (MS-port) (1006). The MS-ports are ~0.9 cm from the edge of the disc (not shown). The disc was of the same size as a conventional CD. The CD has a home mark (1035) at the edge (1036) for
5 positioning the disc when dispensing liquids.

The final device comprises a bottom part in plastic material that contains the uncovered form of the microchannel structures given in **figure 8a**. The microchannel structures are covered with a lid in which there are circular
10 holes (1007,1008,1009,1010,1011,1012 in figure 7b) that will function as inlets (1007,1008) or outlets (1009,1010) in the final microfluidic device or as separate claibrator areas (1011,1012). The bottom part with its microstructures is made of plastics and has been manufactured by a moulding replication process. The surface with the uncovered form of the
15 microchannel structures has been hydrophilised in accordance WO 0056808 (Gyros AB). The lid was thermo laminated to the bottom part in accordance with **WO 0154810** (Gyros AB).

Figure 8b shows in enlarged form a set (1001) of 10 microchannel structures
20 (1002). Each microchannel structure has a sample inlet port (1005) and one common inlet port (1004) for other liquids. At the bottom of each of these two inlet ports (1004,1005) there are ridges/grooves (1013) directed inwards the microchannel structure. The sample inlet port (1005) is connected to one (1014) of two inwardly/upwardly directed shanks (1014,1015) of a Y-shaped
25 sample reservoir (1016). The inlet port (1004) for other liquids is common for all microchannel structures in a set and is connected to a common distribution manifold (1017) with one reservoir/volume defining unit (1018) for other liquids than sample connected to the other upwardly directed shank (1015) of each sample reservoir (1016). The distribution manifold (1017) has one waste
30 outlet port (1009) at each flank of the set. The downward shank (1019) of the Y-like sample reservoir (1016) leads to an outlet port (MS port) (1006) and comprises a bed (1020) of chromatography particles (RPC, reversed phase chromatography) held against a dual depth (1021) (from 100 µm to 10 µm to

20 μm in the flow direction), of the outer part of the downward shank/microchannel (1019). The microchannel corresponding to the downward shank (1021) will end in the bottom (1022) of the outlet port (MS port) (1006) as a widening groove (drop-like seen from above)(1023), which
5 will function as a crystallization area.

Each volume-defining unit (1018) for other liquids is surrounded by anti-wicking means (1024,1025) that will prevent wicking of liquid between the volume-defining units (1018). The anti-wicking comprises both (a) a geometric
10 change (1024) in edges going between the volume defining units (1018) or from a volume defining unit (1018) to a waste outlet port (1009) and a hydrophobic surface break (1025, rectangle).

Valve functions in the form of local hydrophobic surface breaks (rectangles,
15 1026, 1027) are present in the waste channels (1028) of the distribution manifold (1017) before the outlet openings (1009) at the flanks, and in each microconduit (1029) between a volume defining unit (1018) for other liquids and the upwardly directed shank (1015) of the sample reservoir (1016). The valve function (hydrophobic surface break) (1027) may be positioned before,
20 across or immediately after the joint between the microconduit (1029) and the upward shank (1015) of the sample reservoir (1016). Despite the sharp change in lateral dimension at the joint between the microconduit (1029) and the upward shank (1015), the hydrophobic surface break (1027) was imperative for the valve function.

25

Local hydrophobic surface breaks (1030,1031, rectangles) for directing liquid into the structure are present at the inlet openings (1007,1008).

Furthermore, a U- (horse-shoe) shaped local hydrophobic surface break
30 (1032) is positioned at the outlet opening (1010 of each outlet port (1006, MS-port) for preventing liquid exiting into the port from spreading onto the top of the disc.

The hydrophobic surface breaks (1026,1027,1030,1031) were applied before an upper substrate (lid) was laminated to the surface of the bottom substrate comprising the microchannel structures in open form. The hydrophobic surface break (1032) was applied after lamination and gold sputtering.

5

The openings (10011,1012) in the lid are calibration areas for calibration substance. The surface within the circles is the top of the bottom part. One (1012) of them comprises a depression (1033) that mimics the widening groove (1022) of an MS-port (1006)

10

Before application of the local hydrophobic surface area (1032) around the opening (1010) the top of the lid was sputtered with gold at least as a continuous layer in-, around-, and between the openings including the calibrator areas (1010). A continuous gold film thus were connecting the
15 bottom and the walls of the MS-ports (1006) and the calibrator areas . Other parts of the lid (but not the whole lid), besides the areas in and around the MS-ports and calibrators, were also covered with gold. The aim has been to cover as much lid area as possible with gold as long as the gold layer do not interfere with microfluidic- and instrumental functions, e.g. the gold is not
20 allowed to cover the rim of the lid (CD) as it upsets the home-positioning of the CD or the inlets (1007,1008) of the microfluidic structures since it affects the capillary force by an increased hydrophobicity (liquid would then be more difficult to fill up the channels). Other conducting materials than gold could also be beneficial for this application, for instance at the filing date indium tin
25 oxide was sputtered onto the lid and was shown promising for this application. Since indium tin oxide is much more transparent than gold and relatively hydrophilic the whole lid could be covered (i.e. no mask would be necessary for sputtering the conductive layer) without concern for microfluidics and instrumental aspects. Therefore the manufacturing and production process
30 would be more simple and cheap.

The depth in the microchannel structures is the same (100 μm) and constant from the inlet openings (1007,1008) to the dual depths (1021).

Loading of RPC-particles. The distribution manifold (1017) is filled with a suspension of RPC-particles via the common inlet port (1004). After filling, the suspension will be present between the inlet port (1004) and the valves (1026) at the flanking waste openings (1009). Upon spinning at a first speed, excess waste suspension will leave the distribution manifold (1017) via the flank openings (1009) while air will enter the manifold via the common inlet (1004). Defined aliquots (about 0,2 μ l) of the suspension will be retained in the volume-defining units (1018). The anti-wicking means (1024,1025) surrounding the volume-defining units (1018) will assist in retaining the defined volume in each volume-defining unit. When the spinning speed is increased, the aliquots in the volume-defining units (1018) will break through the valves (1027), pass through upward shanks (1015) and the Y-shaped sample reservoirs (1016) and out through the downward shank (1019). The particles will be collected as a packed bed (1020) against the dual depth (1021), and the liquid will pass out through the outlet opening (1012) where it leaves the system.

Filling of the distribution manifold (1017) including the volume defining units (1018) through the common inlet port (1004) is solely by capillary force.

20

Experimental

A model protein consisting of bovine serum albumin (BSA) in 50 mM ammonium bicarbonate buffer, pH 8, was reduced and alkylated according to standard protocol and in-solution digested with trypsin. The reaction was quenched by adding trifluoroacetic acid (TFA) to a final concentration of 0.1% and transferred to a micro plate for subsequent sample processing on-CD, as described above.

Sample and reagents were transferred from micro plates (containing typical volumes of 5 to 100 μ l) to CD by a robotic arm. The robotic arm holds 10 capillaries where sample and reagents are contained inside during transfer. The volume of sample/reagents aspirated into the capillaries and later dispensed onto the CD is driven by syringe pumps and controlled by software

(as are the robotic arm). Aspiration and dispensation rates are typical in the 0.5-10 $\mu\text{l}/\text{sec}$ rate. Once the liquid is dispensed onto the CD, at respective inlet port, it is drawn, by capillary force, into respective common/micro structure.

- 5 The instrument for performing the experiment was a CD microlaboratory (Gyrolab Workstation, Gyros AB, Uppsala, Sweden). This instrument is a fully automated robotic system controlled by application-specific software. Microplates containing samples or reagents are stored in a carousel within the system. A high precision robot transfers samples from microplates or
10 containers into the microworld of the CD. CDs are moved to the spinning station for the addition of samples and reagents. An application-specific method within the software controls the spinning at precisely controlled speeds controls the movement of liquids through the microstructures as the application proceeds. The CDs are transferred to a MALDI mass
15 spectrometer for analysis and identification.

In order to reduce eventual carry-over between individual microchannel structures, i.e., if part of sample remains inside the capillary after dispensing it onto CD it might contaminate the sample following and has therefore to be
20 properly washed away, the following wash procedure was applied:

1. 20 μl of water was flushed through all capillaries.
2. 4 μl of 50% ethanol in water was then aspirated into the capillaries and dispensed to waste, this was repeated four times using 4.5 μl in the last
25 two cycles.
3. Finally, 4 μl of 0.1% TFA was aspirated and dispensed to waste, this was repeated four times using 4.5 μl in the last two cycles.

Operation method:

- 30 The following scheme gives an overview of a typical spin program for running multiplex samples on a CD for the above-mentioned MALDI application. The CD is the one described above. A ramp (see below) indicates an acceleration phase, deceleration phase, or a constant rpm value.

The CD was applied in an instrument from Gyros AB.

1. *First spin.* The purpose here is to restore ("re-pack") the chromatographic columns.

Order	Ramp (rpm)	Time (sec)	Order	Ramp (rpm)	Time (sec)
1	7000	2	3	0	2
2	7000	30			

5

2. *Conditioning of common/individual microstructures and reversed-phase columns.* 3.8 µl (per 10 structures) of 50% acetonitrile in water is dispensed into each common inlet port (1004). The first ramp (no spin) is a lag period as for the liquid to completely fill up the common channel.

Order	Ramp (rpm)	Time (sec)	Order	Ramp (rpm)	Time (sec)
1	0	5	6	0	2
2	700	7	7	8000	2
3	700	2.5	8	8000	30
4	1600	0.15	9	0	2
5	1600	20			

10

3. *Conditioning of individual microstructures.* This item differs from the one above (no 2) by addressing other parts of the microchannel structures not accessible by the procedure mentioned in item 2. The purpose is to more completely re-wett any microstructure. 400 nl of 50% acetonitrile in water is dispensed per microchannel structure through each inlet ports (1005).

Order	Ramp (rpm)	Time (sec)	Order	Ramp (rpm)	Time (sec)
1	8000	2	3	0	2
2	8000	30			

4. *Conditioning of common/individual microstructures and reversed-phase columns.* 3.8 µl (per 10 structures) of 0.1% trifluoroacetic acid (TFA) in water

is dispensed into each common inlet port (1004). The first ramp (no spin) is a lag period as for the liquid to completely fill up the common channel.

Order	Ramp (rpm)	Time (sec)	Order	Ramp (rpm)	Time (sec)
1	0	5	5	1600	20
2	700	7	6	8000	2
3	700	2.5	7	8000	30
4	1600	0.15	8	0	2

5. *Sample transfer.* 1-10 μ l of sample is applied into each inlet port (1005) (total 100 identical micro structures and therefore 100 samples per CD). The first ramp (no spin) is a lag period as for the liquid to completely fill up the common channel.

Order	Ramp (rpm)	Time (sec)	Order	Ramp (rpm)	Time (sec)
1	0	5	7	1200	20
2	1800	0.3	8	2500	0.25
3	1000	0.2	9	1500	0.2
4	1000	30	10	1500	20
5	2000	0.2	11	0	2
6	1200	0.2			

6. *Desalting/washing of sample.* 3.8 μ l (per 10 structures) of 5-10% organic solvent/ 0.1% trifluoroacetic acid (TFA) in water is dispensed into each common inlet port (1004). The first ramp (no spin) is a lag period as for the liquid to completely fill up the common channel.

Order	Ramp (rpm)	Time (sec)	Order	Ramp (rpm)	Time (sec)
1	0	5	5	1600	20
2	700	7	6	8000	2
3	700	2.5	7	8000	30
4	1600	0.15	8	0	2

7. Sample elution and peptide-matrix cocrystallization on MALDI target area on CD.

Eluent consists of 50% acetonitrile/0.1% TFA in water wherein the MALDI matrix (1.5 mg/ml of α -cyanohydroxycinnamic acid) is dissolved. 4.1 μ l of
5 eluent (per 10 structure) is applied into each common inlet port (1004).

Order	Ramp (rpm)	Time (sec)	Order	Ramp (rpm)	Time (sec)
1	0	2	15	300	4
2	600	0.1	16	1600	0.2
3	600	7	17	1600	0.1
4	1400	0.14	18	1200	0.07
5	1400	0.25	19	1200	0.4
6	300	0.22	20	1000	0.05
7	300	4	21	1000	1
8	1400	0.2	22	800	0.05
9	1400	0.1	23	800	90
10	300	0.2	24	1200	0.1
11	300	4	25	1200	1.9
12	1400	0.2		800	0.1
13	1400	0.1		800	90
14	300	0.1		0	2

The CD (or more exactly half of it) was subsequently fixed to a steel target holder and inserted into a MALDI TOF instrument (Bruker Biflex) for running
10 mass spectrometry.

Results:

The molecular mass of the peaks was identified as BSA peptides by a database search (NCBI). The mass spectra typically showed ten peaks which
15 were identified as BSA peptides. High sensitivity was attainable using the CD for sample concentration and preparation. High mass resolution and accuracy were also demonstrated.

Comments on the design of the MS-port

20 Meanwhile the peptides are eluted from the chromatographic column with an organic:aqueous solvent containing the MALDI matrix, the liquid flows into the MS-port (i.e., the MALDI target area) by centrifugal force. Once a liquid

element (droplet) enters this open area (restricted by the walls of the lid and the upper surface of the bottom substrate) the solvent quickly evaporates and peptides and matrix cocrystallizes on the surface. In order to make this process more robust, i.e., to stronger retain the liquid element while spinning

5 is performed, a hydrophobic pattern was created surrounding the MS-port (then considered a more hydrophilic area). This process of hydrophobic patterning and its flow restriction effect is similar to the process and effect of creating hydrophobic breaks, the difference here being that the hydrophobic pattern surrounding the MS-port is created after the lid has been laminated

10 onto the CD and after the application of the conductive layer. This hydrophobic area has a U-shape (horseshoe) configuration and covers part of the MS-port and part of the lid surface and its wall. Since the liquid element is repelled from this hydrophobic area the droplet preferably stays on the more hydrophilic area during crystallization. In addition to this the crystals are

15 formed on a smaller surface area at some distance away from the walls of the lid. This means that the analyte concentration will be further enhanced (and therefore a higher sensitivity can potentially be reached in the subsequent mass spectrometry analysis) compared to if the crystals were deposited on a larger surface where the sample would more spread out. Also, with

20 automated MALDI analysis it is preferable to have a smaller surface area where the crystals are found as for the laser to more efficiently cover that particular area in a shorter time period (assuming heterogeneous crystal formation, i.e., no "sweet spot"). Moreover, by having the crystals at some distance away from the lid wall less electric field strength disturbances are

25 expected during MALDI analysis due to a non-homogenous field close to the wall. If so, less mass accuracy and resolution is expected. The same would be true for crystals found at different height levels attached to the wall of the lid, i.e., less mass accuracy and resolution would be expected if the crystals were to be irradiated by the laser at different heights along the wall. Finally,

30 any influence of "laser-shadow" by the wall will be diminished.

Certain innovative aspects of the invention are defined in more detail in the appending claims. Although the present invention and its advantages have

been described in detail, it should be understood that various changes, substitutions and alterations can be made herein without departing from the spirit and scope of the invention as defined by the appended claims.

Moreover, the scope of the present application is not intended to be limited to
5 the particular embodiments of the process, machine, manufacture, composition of matter, means, methods and steps described in the specification. As one of ordinary skill in the art will readily appreciate from the disclosure of the present invention, processes, machines, manufacture, compositions of matter, means, methods, or steps, presently existing or later
10 to be developed that perform substantially the same function or achieve substantially the same result as the corresponding embodiments described herein may be utilized according to the present invention. Accordingly, the appended claims are intended to include within their scope such processes, machines, manufacture, compositions of matter, means, methods, or steps.

15

C L A I M S

1. A method for presenting an analyte of a liquid sample as an MS-analyte to a mass spectrometer, **characterized** in
 - (a) comprising the steps of:
 - 5 (i) applying the liquid sample to a sample inlet port (I) of a microchannel structure (I) of a microfluidic device, said structure also comprising an MS-port,
 - (ii) transporting the analyte by a liquid flow in microchannel structure (I) thereby transforming the analyte to an MS-analyte, and
 - 10 (iii) presenting the MS-analyte to a mass spectrometer via the MS-port, and
 - (b) using inertia force for creating said liquid flow within at least a part of microchannel structure (I).
- 15 2. The method of any of claims 1, **characterized** in that
 - (a) the device comprises a disc with an axis of symmetry perpendicular to the disc,
 - (b) microchannel structure (I) is oriented radially with a liquid flow direction from an inner inlet port towards the periphery of the disc, and
 - 20 (c) inertia force is centrifugal force which is created by spinning said substrate around the axis of symmetry.
3. The method of claim 2, **characterized** in that centrifugal force is used for driving liquid into the MS-port.
- 25 4. The method of any of claims 1-3, **characterized** in that microchannel structure (I) comprises a separation zone with a separation medium between said sample inlet port and the MS-port and that step (ii) comprises that
 - 30 (a) said analyte or an analyte-derived entity is bound to said separation medium, and thereafter
 - (b) a liquid that releases said analyte or the analyte-derived entity from

said separation medium for transport to the MS-port is introduced into microchannel structure (I) upstream said separation zone.

5. The method of claim 4, **characterized** in that microchannel structure (I) comprises an inlet port (II) which is separate from inlet port (I) and that said liquid for release of the analyte or the analyte-derived entity from the separation medium is introduced via inlet port (II).
6. The method of any of claims 2-5, **characterized** in that the MS-port is
 - (a) downstream said inner inlet port which is equal to inlet port (I) and
 - (b) located at a larger radial distance from the axis of symmetry than said inner inlet port.
7. The method of any of claims 2-6, **characterized** in that
 - (a) said disc comprises two or more microchannel structure (I) which are annularly arranged around the axis of symmetry with MS-ports being located at essentially the same radial distance from the axis of symmetry, and
 - (b) steps (i) – (iii) are applied to at least one microchannel structure (I).
8. The method of any of claims 1-7, **characterized** in that the MS-port in microchannel structure (I) comprises an electrospray arrangement.
9. The method of any of claims 1-7, **characterized** in that the MS-port in microchannel structure (I) comprises an EDI-area comprising a conductive layer (I) with a conductive connection.
10. The method of claim 9, **characterized** in that step (ii) comprises that
 - (a) an EDI matrix is included in a volatile liquid which is introduced into at least one microchannel structure (I),
 - (b) the volatile liquid, the EDI-matrix, and the MS-analyte are allowed to enter simultaneously each MS port of said at least one microchannel structure (I) by the application of centrifugal force,

(c) evaporating the volatile liquid under the application of centrifugal force while cocrystallizing the EDI-matrix with the MS-analyte.

11. A microfluidic disc comprising
- 5 (a) an axis of symmetry perpendicular to the plane of the disc,
(b) a microchannel structure (I) comprising an inner application area at a shorter radial distance than an outlet port and comprising an MS-port and a sample inlet port (I).
- 10 12. The disc of claim 11, **characterized** in that microchannel structure (I) comprises a reaction zone between said sample inlet port and the MS-port.
13. The disc of claim 12, **characterized** in that said reaction zone is a
- 15 separation zone comprising a separation medium which is capable of binding an analyte.
14. The disc of any of claims 11-13, **characterized** in that microchannel structure (I) comprises an inlet port (II) for introduction of a liquid other
- 20 than the sample into microchannel structure (I).
15. The disc of claim 14, **characterized** in that inlet port (I) and inlet port (II) coincides or are separate.
- 25 16. The disc of any of claims 11-15, **characterized** in that said MS-port is
(a) downstream said inner inlet port which is equal to inlet port (I) and
(b) located at a larger radial distance from the axis of symmetry central axis than said inner inlet port.
- 30 17. The disc of any claims 11-16, **characterized** in that said disc comprises two or more microchannel structure (I) which are annularly arranged around the axis of symmetry with their MS-ports being located at essentially the same radial distance from the axis of symmetry.

18. The disc of any of claims 11-17, **characterized** in that the MS-port in microchannel structure (I) comprises an electrospray arrangement.
19. The disc of any of claims 11-17, **characterized** in that the MS-port in
5 microchannel structure (I) is and EDI MS-port comprising an EDI-area with a conductive layer (I).
20. The disc of claim 19, **characterized** in that said EDI-area is an LDI-area.
- 10 21. The disc of any of claims 19-20, **characterized** in that layer (I) comprises a conductive metal.
22. The disc of any of claims 19-20, **characterized** in that layer (I) comprises a conductive metal oxide.
- 15 23. The disc of any of claims 19-22, **characterized** in that the disc comprises
(a) two or more microchannel structure (I),
(a) a continuous conductive layer which comprises layer (I) of each MS-port of said two or more microchannel structures.
- 20 24. The disc of any of claims 19-23, **characterized** in that the layer (I) is exposed as an EDI-surface in the MS-port of microchannel structure (I).
- 25 25. The disc of any of claims 23-24, **characterized** in that said continuous conductive layer is exposed on the surface of the disc.
26. The disc of any of claims 19-25, **characterized** in that layer (I) has a conductive connection.
- 30 27. The disc of claim 26, **characterized** in that layer (I) is part of a continuous conductive layer and that this layer provide the conductive connection.

28. The disc of any of claims 26-27, **characterized** in that there is a calibrator area associated with each MS-port, each calibrator area possibly being common for two or more MS-ports.

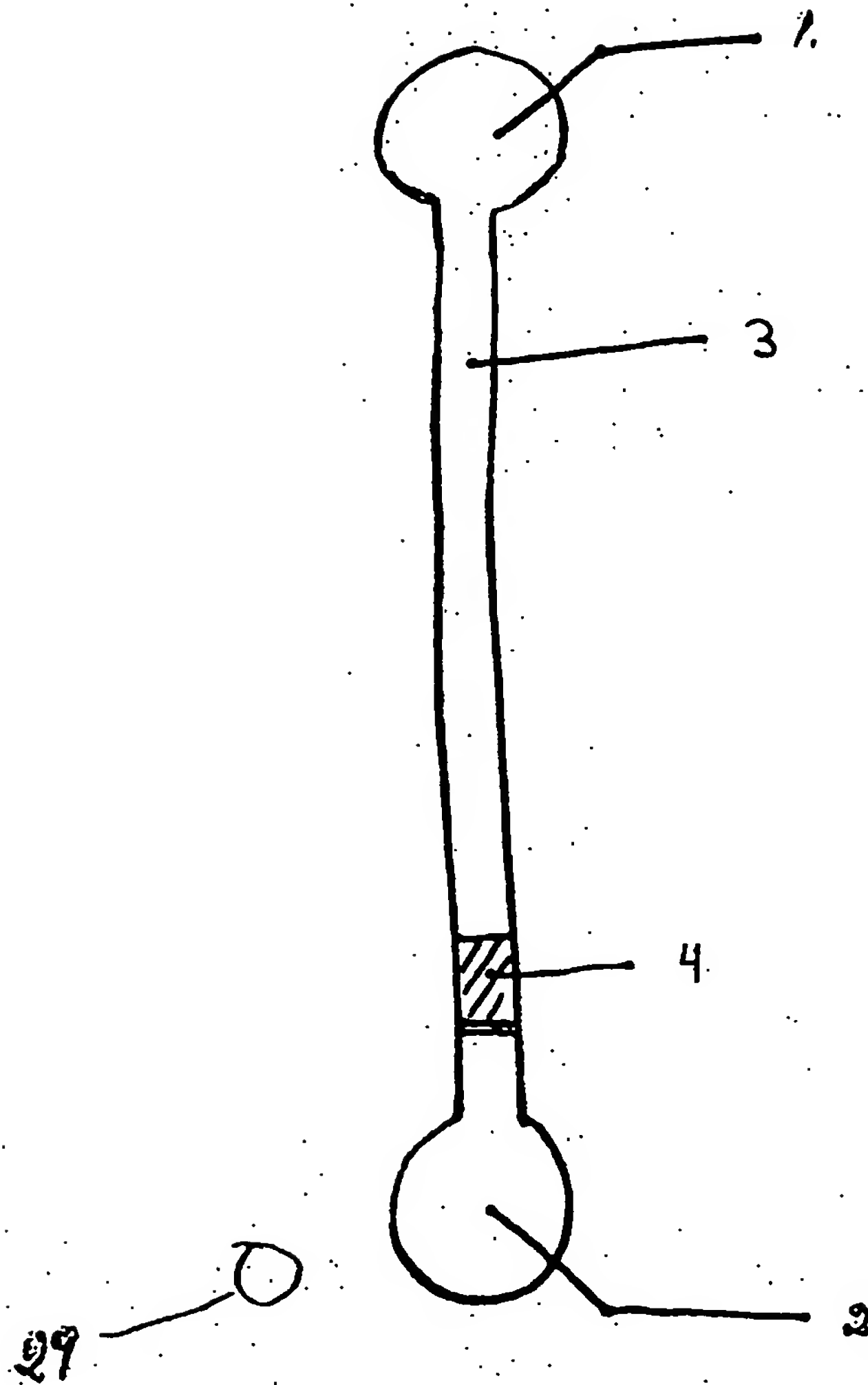


Fig. 1

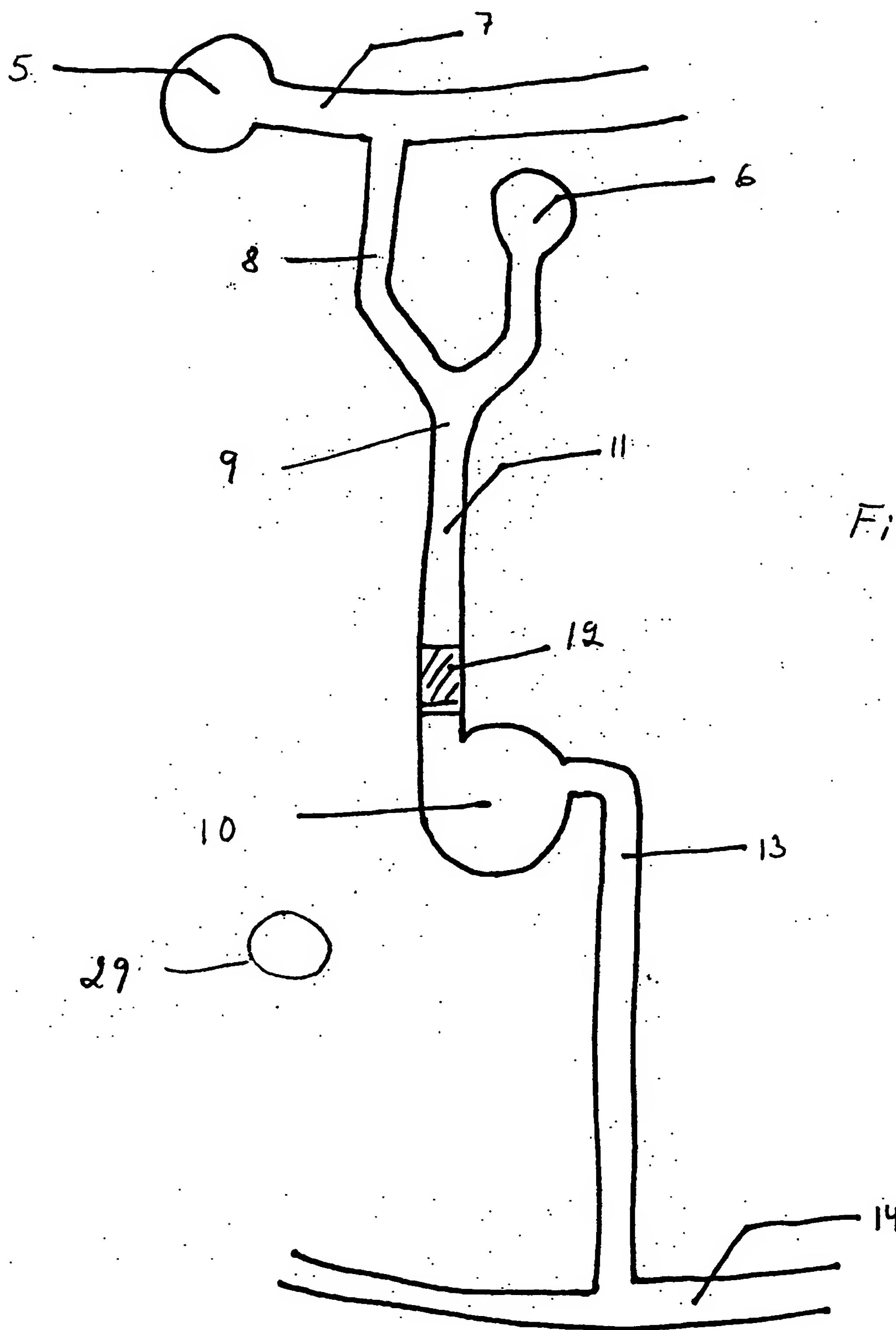
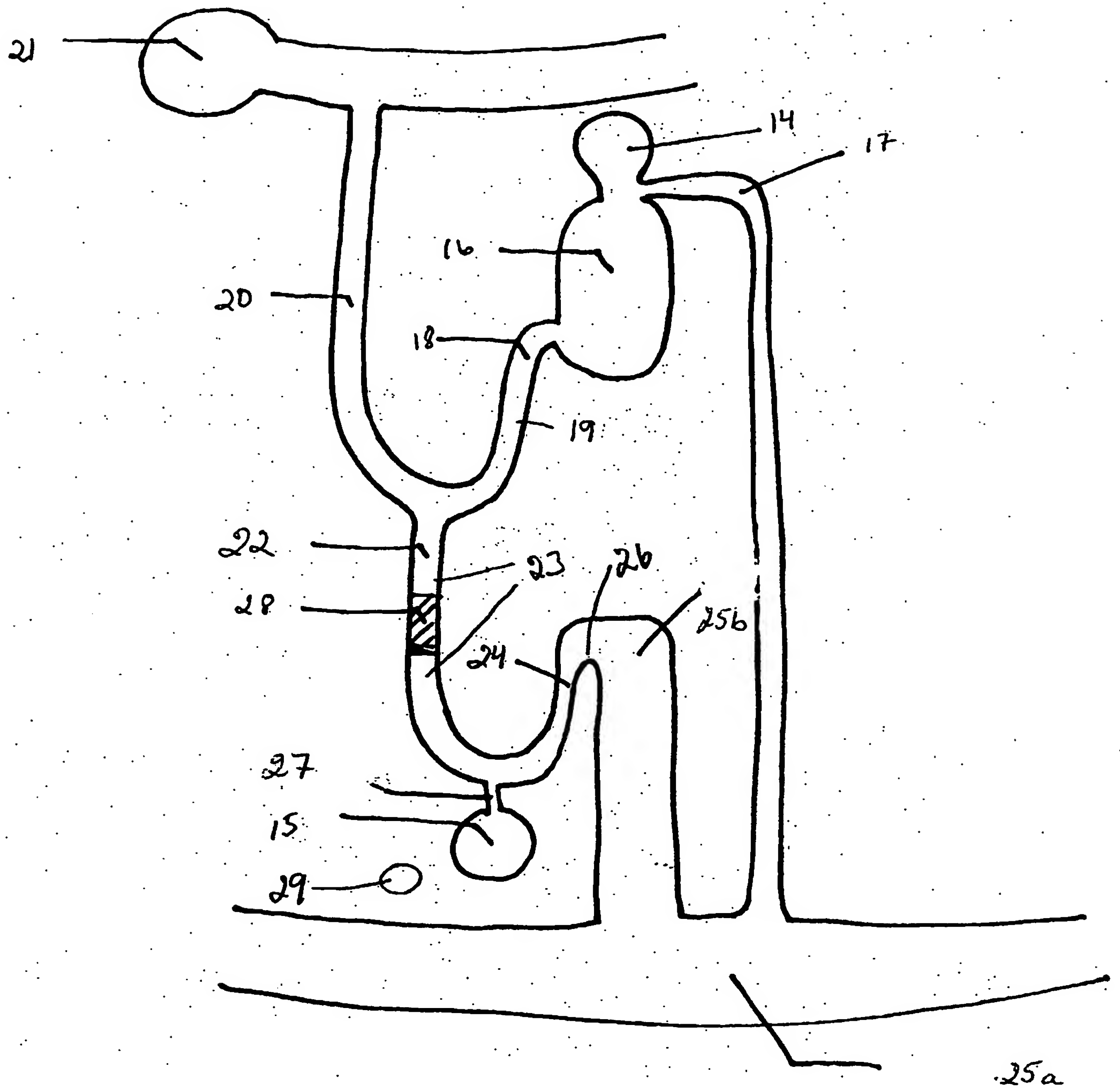


Fig 2



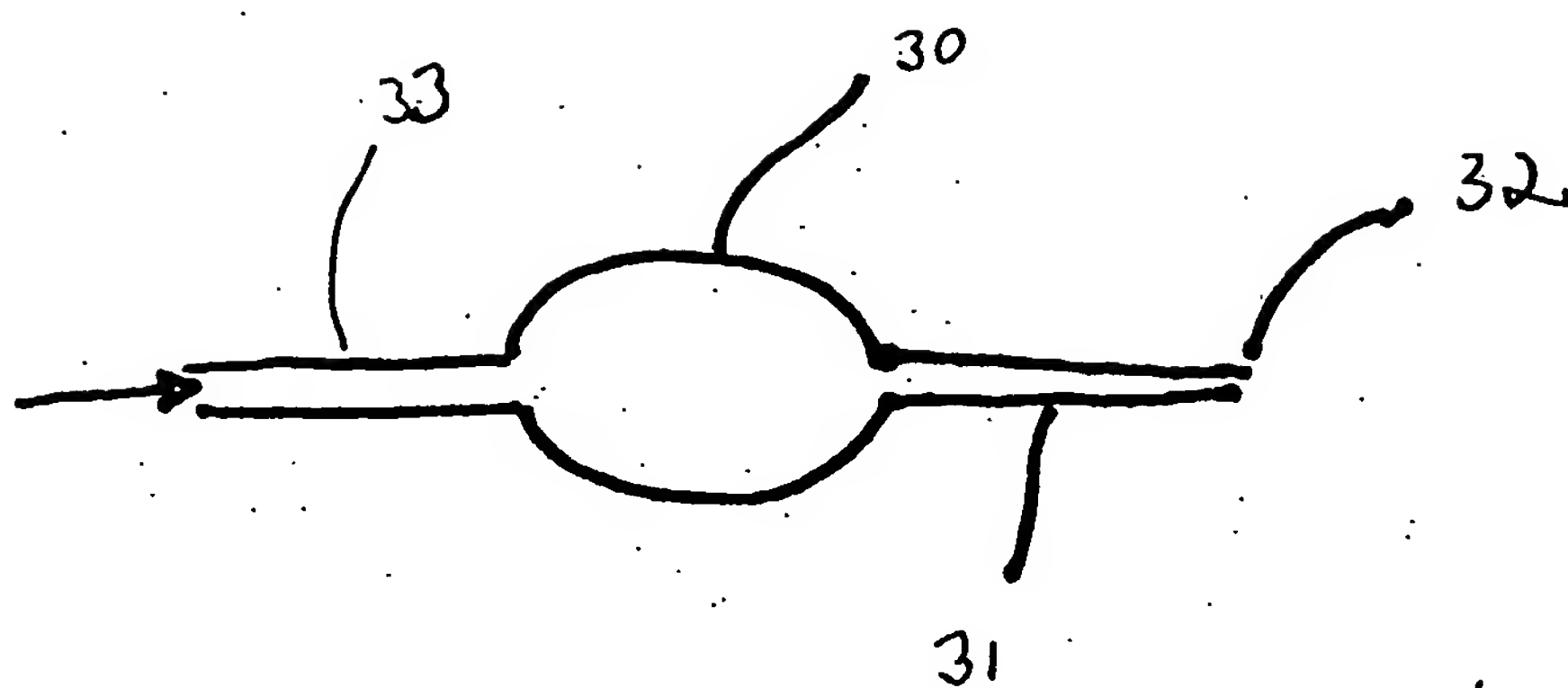
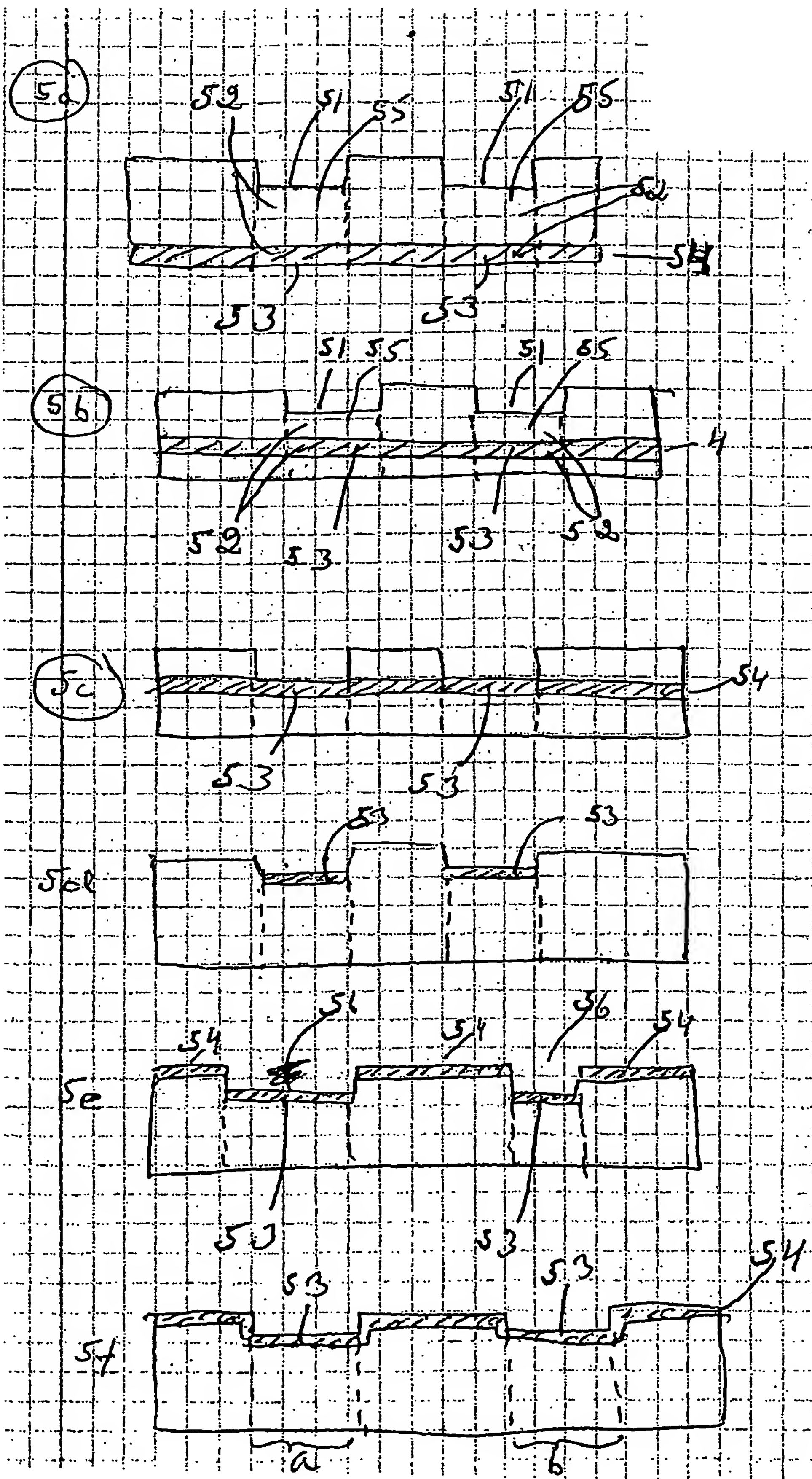


Fig 4



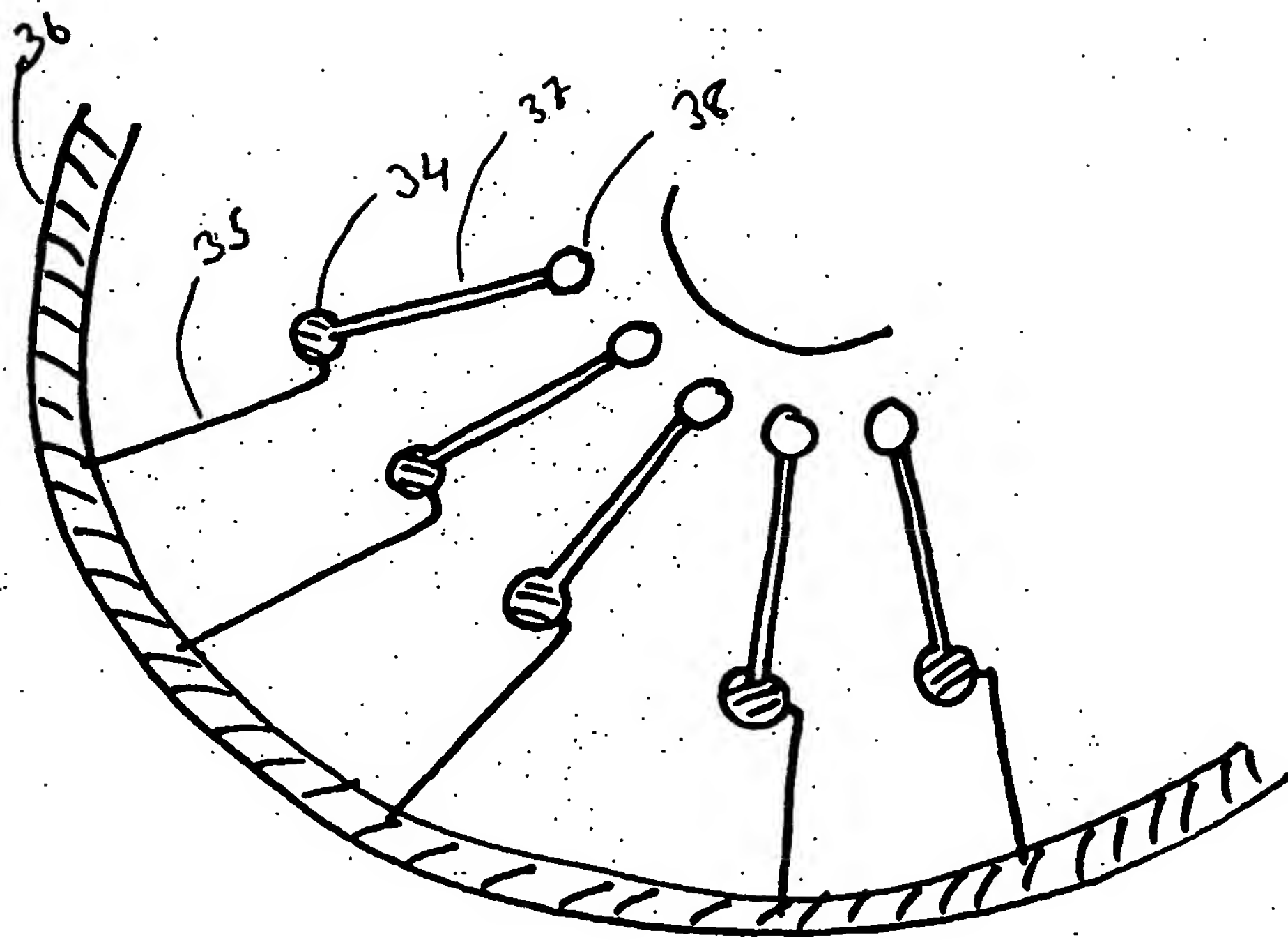


Fig 6

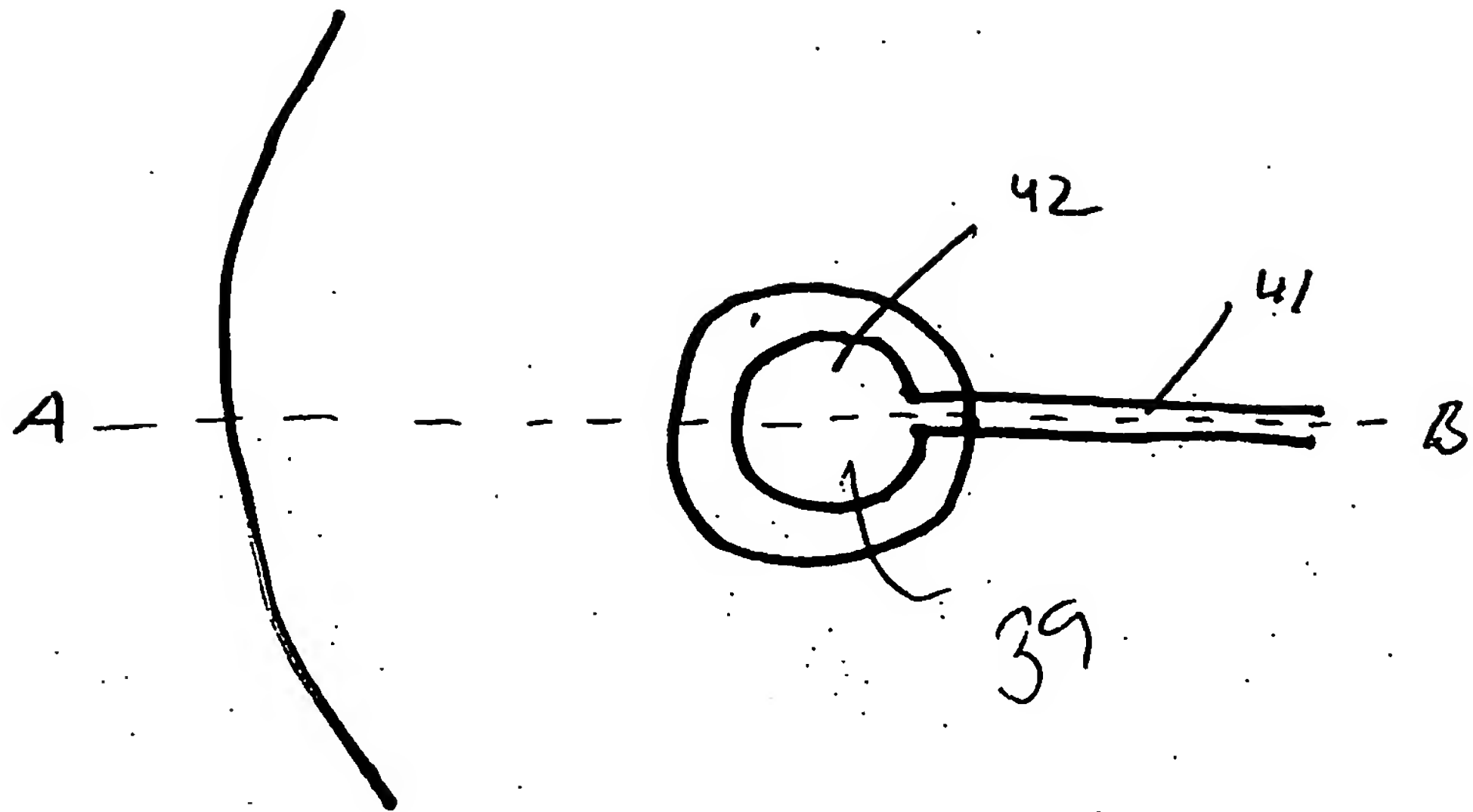


Fig 7a

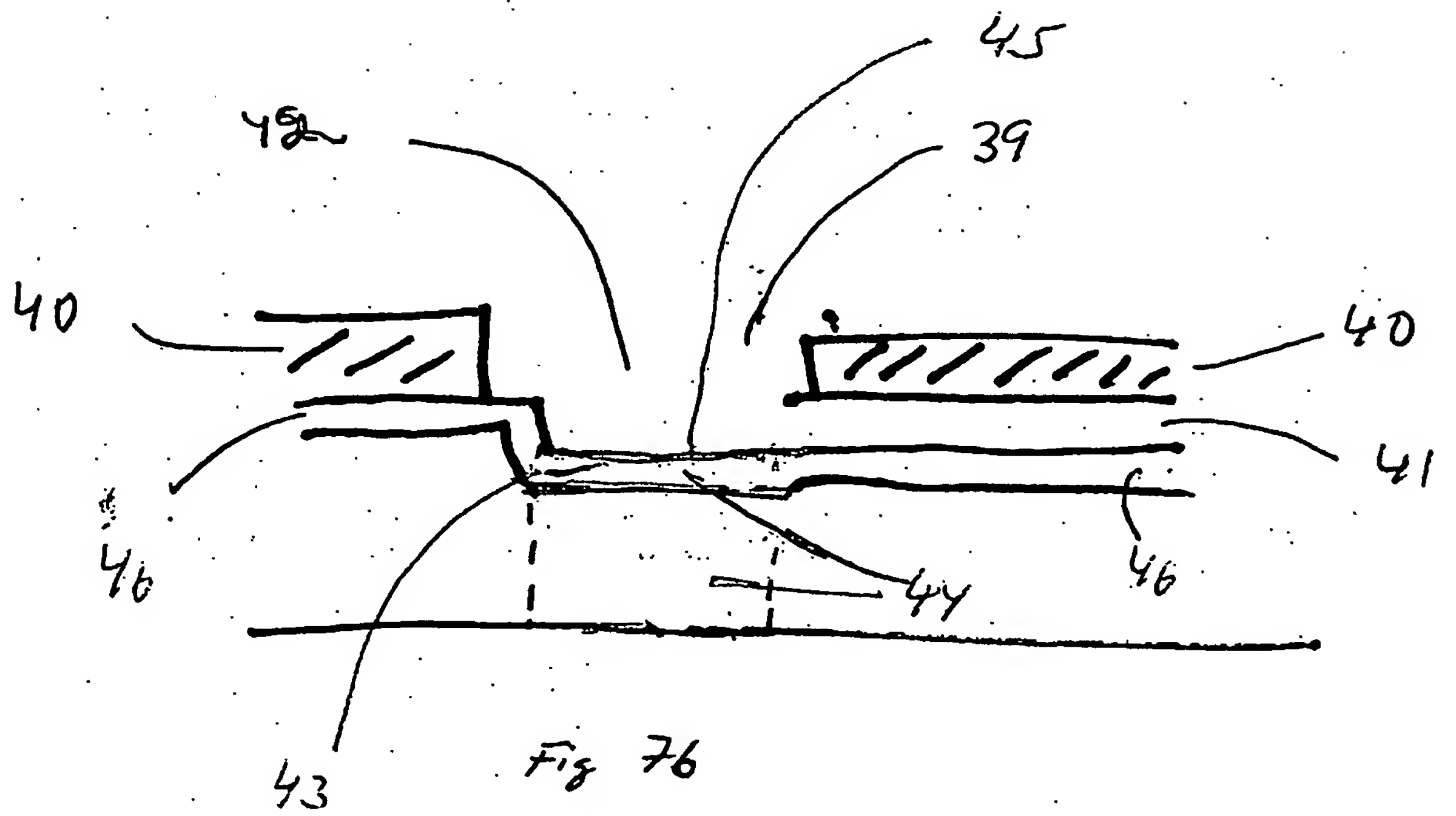


Fig 7b

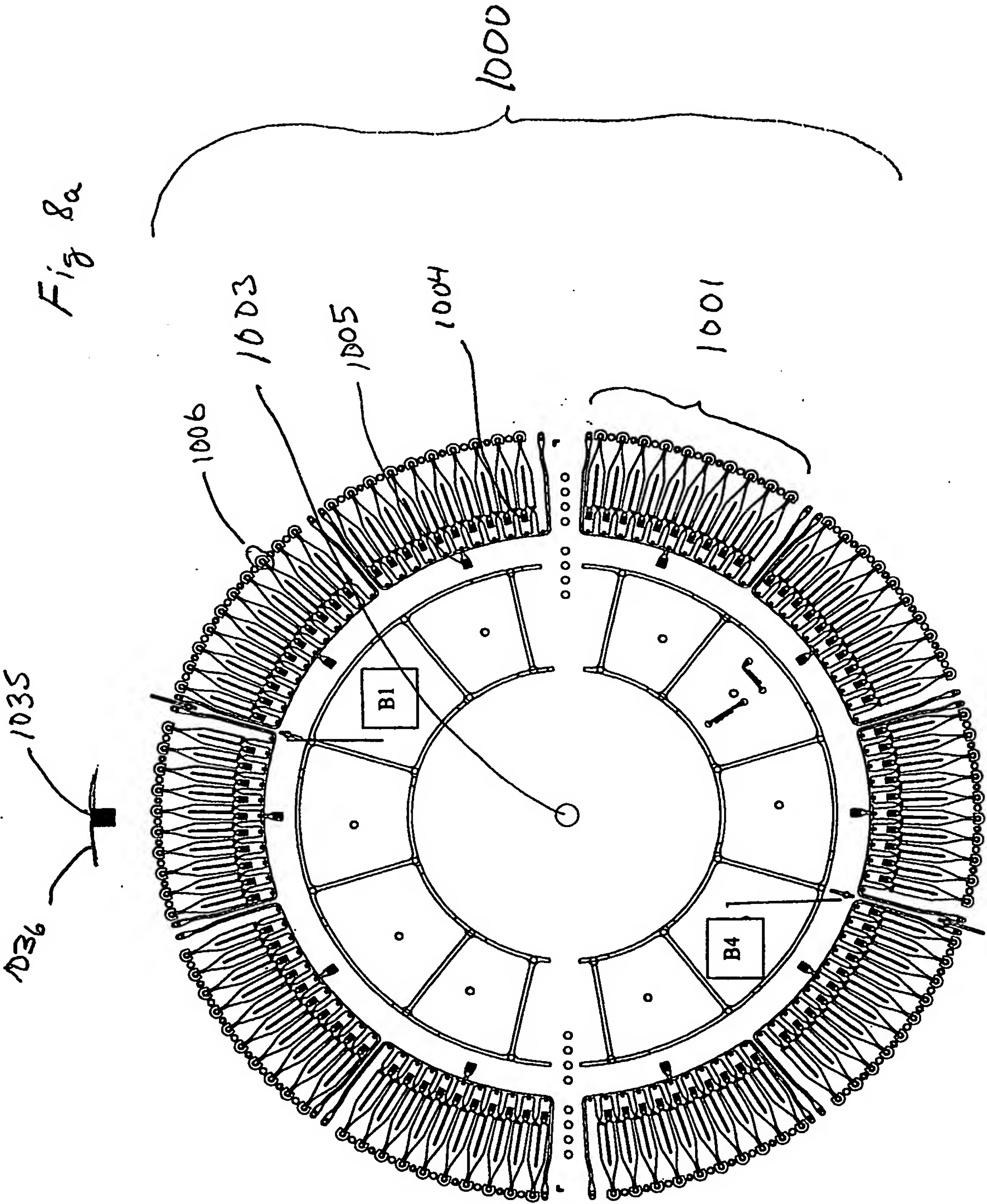
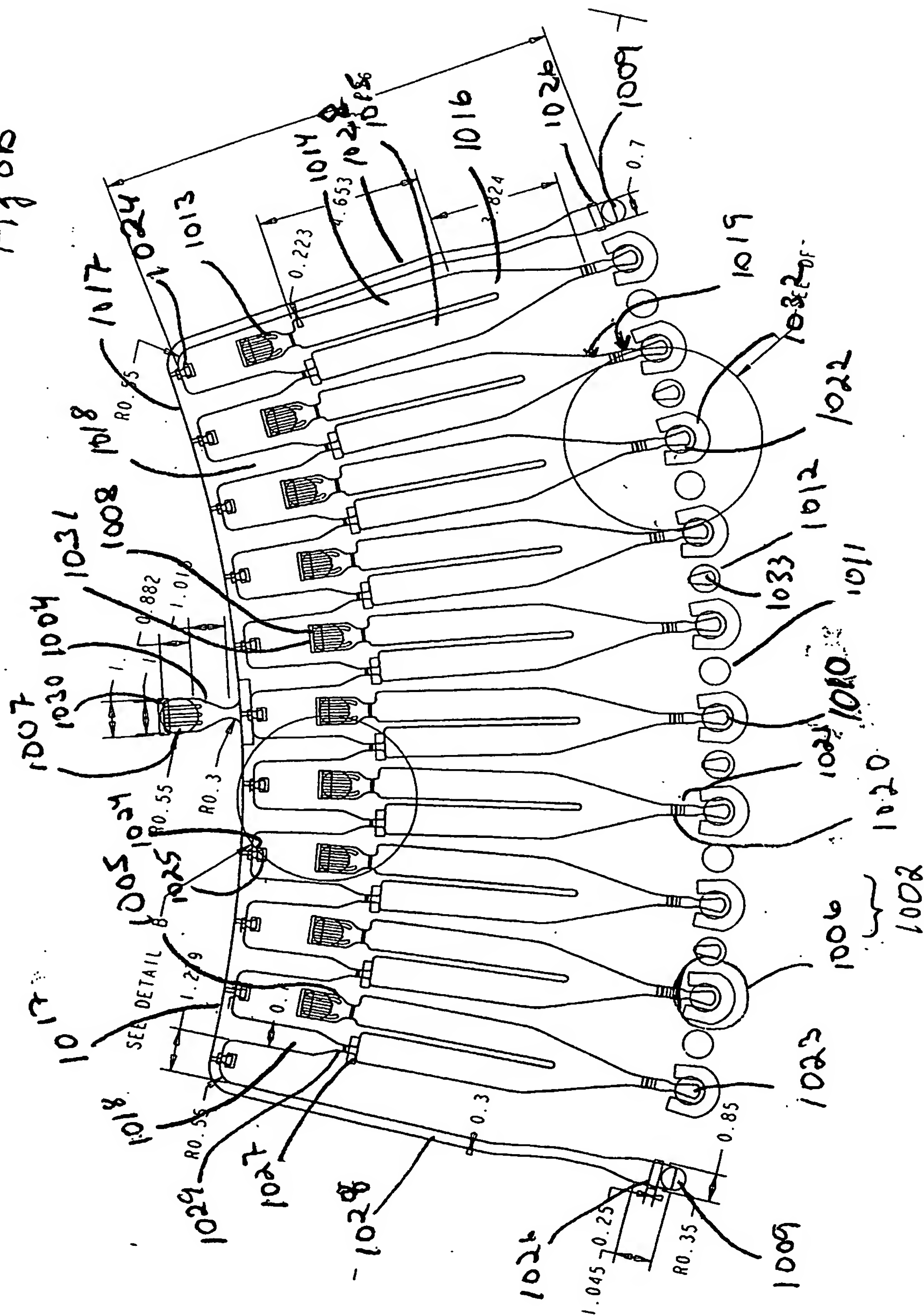


Fig 8b



INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 02/00539

A. CLASSIFICATION OF SUBJECT MATTER

IPC7: H01J 49/04, G01N 35/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC7: G01N, H01J

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-INTERNAL, WPI DATA, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 0030167 A1 (CALIFORNIA INSTITUTE OF TECHNOLOGY), 25 May 2000 (25.05.00), see whole document	1-3,6-12, 14-28
A	--	4-5,13
Y	WO 9721090 A1 (GAMERA BIOSCIENCE), 12 June 1997 (12.06.97), abstract	1-3,6-12, 14-28
A	WO 0067293 A1 (CIPHERGEN BIOSYSTEMS, INC.), 9 November 2000 (09.11.00), see whole document	1-28



Further documents are listed in the continuation of Box C.



See patent family annex.

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- "&" document member of the same patent family

Date of the actual completion of the international search

26 June 2002

Date of mailing of the international search report

28-06-2002

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INTERNATIONAL SEARCH REPORT

Information on patent family members

10/06/02

International application No.

PCT/SE 02/00539

Patent document cited in search report			Publication date	Patent family member(s)		Publication date
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